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**UTILITY
PATENT APPLICATION
TRANSMITTAL**

(Only for new nonprovisional applications under 37 CFR 1.53(b))

Attorney Docket No.	7991-086	Total Pages	63
<i>First Named Inventor or Application Identifier</i>			
Beetham et al.			
Express Mail Label No.	EL 501 634 414 US		

APPLICATION ELEMENTS

See MPEP chapter 600 concerning utility patent application contents.

1. Fee Transmittal Form
Submit an original, and a duplicate for fee processing
2. Specification including Table of Contents
(preferred arrangement set forth below)
 - Descriptive title of the Invention
 - Cross Reference to Related Applications
 - Statement Regarding Fed sponsored R&D
 - Reference to Microfiche Appendix
 - Background of the Invention
 - Brief Summary of the Invention
 - Brief Description of the Drawings *(if filed)*
 - Detailed Description of the Invention (including drawings, *if filed*)
 - Claim(s)
 - Abstract of the Disclosure
3. Drawing(s) (35 USC 113) [Total Sheets 10]
4. Oath or Declaration [Total Sheets 2]
 - a. Newly executed (original or copy)
 - b. Copy from a prior application (37 CFR 1.63(d))
(for continuation/divisional with Box 17 completed)
[Note Box 5 below]
 - i. DELETION OF INVENTOR(S)
Signed statement attached deleting inventor(s) named in the prior application, see 37 CFR 1.63(d)(2) and 1.33 (b).
5. Incorporation By Reference *(useable if Box 4b is checked)*
The entire disclosure of the prior application, from which a copy of the oath or declaration is supplied under Box 4b, is considered as being part of the disclosure of the accompanying application and is hereby incorporated by reference therein.

Assistant Commissioner for Patents
ADDRESS TO: Box Patent Application
Washington, DC 20231

6. Microfiche Computer Program *(Appendix)*
7. Nucleotide and/or Amino Acid Sequence Submission
(if applicable, all necessary)
 - a. Computer Readable Copy
 - b. Paper Copy *(identical to computer copy)*
 - c. Statement verifying identity of above copies

10/10/00
10926 US 685403

ACCOMPANYING APPLICATION PARTS

8. Assignment Papers (cover sheet & document(s))
9. 37 CFR 3.73(b) Statement Power of Attorney *(when there is an assignee)*
10. English Translation Document *(if applicable)*
11. Information Disclosure Statement (IDS)/PTO-1449 Copies of IDS Citations
12. Preliminary Amendment
13. Return Receipt Postcard (MPEP 503)
(Should be specifically itemized)
14. Small Entity Statement filed in prior application, Statement(s) Status still proper and desired
15. Certified Copy of Priority Document(s)
(if foreign priority is claimed)
16. Other:

17. If a CONTINUING APPLICATION, check appropriate box and supply the requisite information:
 Continuation Divisional Continuation-in-part (CIP) of prior application No: filed.

18. CORRESPONDENCE ADDRESS

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Date: October 10, 2000

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Assistant Commissioner for Patents
Box PATENT APPLICATION
Washington, D.C. 20231

Sir:

The following utility patent application is enclosed for filing:

Applicant(s): Beetham et al.

Executed on: unexecuted

Title of Invention: NON-TRANSGENIC HERBICIDE RESISTANT PLANTS

PATENT APPLICATION FEE VALUE

TYPE	NO. FILED	LESS	EXTRA	EXTRA RATE	Fee
Total Claims	38	-20	18	\$18.00 each	\$ 324.00
Independent	5	-3	2	\$80.00 each	\$ 160.00
Minimum Fee					\$ 710.00
Multiple Dependency Fee If Applicable (\$270.00)					\$ 270.00
Total					\$ 1,464.00
50% Reduction for Independent Inventor, Nonprofit Organization or Small Business Concern (a verified statement as to the applicant's status is attached)					- \$ 0.00
Total Filing Fee					\$ 1,464.00

This application claims priority to U.S. Provisional Application No. 60/158,027, filed on October 7, 1999 and
U.S. Provisional Application No. 60/173,564, filed December 30, 1999.

Please charge the required fee to Pennie & Edmonds LLP Deposit Account No. 16-1150. A copy of this sheet is
enclosed.

Respectfully submitted,

Thomas E. Friebel

Thomas E. Friebel
PENNIE & EDMONDS LLP

by Thomas E. Friebel

REC'D NO.
40,203 29,258

(Reg. No.)

Enclosure

This form is not for use with continuation, divisional, re-issue, design or plant patent applications.

NON-TRANSGENIC HERBICIDE
RESISTANT PLANTS

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NON-TRANSGENIC HERBICIDE RESISTANT PLANTS

The present application claims priority to U.S. Provisional Application No. 60/158,027, filed on October 7, 1999 and to U.S. Provisional Application No. 60/173,564, 5 filed December 30, 1999, the disclosures of each of which are incorporated by reference herein in their entirety.

1. FIELD OF THE INVENTION

The present invention relates to the production of a non-transgenic plant 10 resistant or tolerant to a herbicide of the phosphonomethylglycine family, *e.g.*, glyphosate. The present invention also relates to the use of a recombinogenic oligonucleobase to make a desired mutation in the chromosomal or episomal sequences of a plant in the gene encoding for 5-enol pyruvylshikimate-3-phosphate synthase (EPSPS). The mutated protein, which substantially maintains the catalytic activity of the wild-type protein, allows for increased 15 resistance or tolerance of the plant to a herbicide of the phosphonomethylglycine family, and allows for the substantially normal growth or development of the plant, its organs, tissues or cells as compared to the wild-type plant irrespective of the presence or absence of the herbicide. The present invention also relates to a non-transgenic plant cell in which the EPSPS gene has been mutated, a non-transgenic plant regenerated therefrom, as well as a 20 plant resulting from a cross using a regenerated non-transgenic plant having a mutated EPSPS gene.

2. BACKGROUND TO THE INVENTION

2.1 PHOSPHONOMETHYLGlyCINE HERBICIDES

25 Herbicide-tolerant plants may reduce the need for tillage to control weeds thereby effectively reducing soil erosion. One herbicide which is the subject of much investigation in this regard is N-phosphonomethylglycine, commonly referred to as glyphosate. Glyphosate inhibits the shikimic acid pathway which leads to the biosynthesis of aromatic compounds including amino acids, hormones and vitamins. Specifically, 30 glyphosate curbs the conversion of phosphoenolpyruvic acid (PEP) and 3-phosphoshikimic acid to 5-enolpyruvyl-3-phosphoshikimic acid by inhibiting the enzyme 5-enolpyruvylshikimate-3-phosphate synthase (hereinafter referred to as EPSP synthase or EPSPS). For purposes of the present invention, the term "glyphosate" includes any herbicidally effective form of N-phosphonomethylglycine (including any salt thereof), other 35 forms which result in the production of the glyphosate anion in plants and any other herbicides of the phosphonomethylglycine family.

Tolerance of plants to glyphosate can be increased by introducing a mutant EPSPS gene having an alteration in the EPSPS amino acid coding sequence into the genome of the plant. Examples of some of the mutations in the EPSPS gene for inducing glyphosate tolerance are described in the following patents: U.S. Patent No. 5,310,667; U.S. Patent No. 5,866,775; U.S. Patent No. 5,312,910; U.S. Patent No. 5,145,783. These proposed mutations typically have a higher K_i for glyphosate than the wild-type EPSPS enzyme which confers the glyphosate-tolerant phenotype, but these variants are also characterized by a high K_m for PEP which makes the enzyme kinetically less efficient (Kishore et al., 1988, Ann. Rev. Biochem. 57:627-663; Schulz et al., 1984, Arch. Microbiol. 137: 121-123; Sost et al., 1984, FEBS Lett. 173: 238-241; Kishore et al., 1986, Fed. Proc. 45: 1506; Sost and Amrhein, 1990, Arch. Biochem. Biophys. 282: 433-436). Many mutations of the EPSPS gene are chosen so as to produce an EPSPS enzyme that is resistant to herbicides, but unfortunately, the EPSPS enzyme produced by the mutated EPSPS gene has a significantly lower enzymatic activity than the wild-type EPSPS. For example, the apparent K_m for PEP and the apparent K_i for glyphosate for the wild-type EPSPS from *E. coli* are 10 μ M and 0.5 μ M, while for a glyphosate-tolerant isolate having a single amino acid substitution of alanine for glycine at position 96, these values are 220 μ M and 4.0 mM, respectively. A number of glyphosate-tolerant EPSPS genes have been constructed by mutagenesis. Again, the glyphosate-tolerant EPSPS had lower catalytic efficiency (V_{max} / K_m), as shown by an increase in the K_m for PEP, and a slight reduction of the V_{max} of the wild-type plant enzyme (Kishore et al., 1988, Ann. Rev. Biochem. 57:627-663).

Since the kinetic constants of the variant enzymes are impaired with respect to PEP, it has been proposed that high levels of overproduction of the variant enzyme, 40-80 fold, would be required to maintain normal catalytic activity in plants in the presence of glyphosate (Kishore et al., 1988, Ann. Rev. Biochem. 57:627-663). It has been shown that glyphosate-tolerant plants can be produced by inserting into the genome of the plant the capacity to produce a higher level of EPSP synthase in the chloroplast of the cell (Shah et al., 1986, Science 233, 478-481), which enzyme is preferably glyphosate-tolerant (Kishore et al., 1988, Ann. Rev. Biochem. 57:627-663).

The introduction of the exogenous mutant EPSPS genes into plant is well documented. For example, according to U.S. Patent No. 4,545,060, to increase a plant's resistance to glyphosate, a gene coding for an EPSPS variant having at least one mutation that renders the enzyme more resistant to its competitive inhibitor, *i.e.*, glyphosate, is introduced into the plant genome. However, many complications and problems are associated with these examples. Many such mutations result in low expression of the mutated EPSPS gene product or result in an EPSPS gene product with significantly lower

enzymatic activity as compared to wild type. The low expression or low enzymatic activity of the mutated enzyme results in abnormally low levels of growth and development of the plant.

While such variants in the EPSP synthases have proved useful in obtaining 5 transgenic plants tolerant to glyphosate, it would be increasingly beneficial to obtain a variant EPSPS gene product that is highly glyphosate-tolerant but still kinetically efficient, such that improved tolerance can be obtained with a wild-type expression level.

2.2 RECOMBINAGENIC OLIGONUCLEOBASES

10 Recombinagenic oligonucleobases and their use to effect genetic changes in eukaryotic cells are described in United States patent No. 5,565,350 to Kmiec (Kmiec I). Kmiec I teaches a method for introducing specific genetic alterations into a target gene. Kmiec I discloses, *inter alia*, recombinagenic oligonucleobases having two strands, in which a first strand contains two segments of at least 8 RNA-like nucleotides that are 15 separated by a third segment of from 4 to about 50 DNA-like nucleotides, termed an “interposed DNA segment.” The nucleotides of the first strand are base paired to DNA-like nucleotides of a second strand. The first and second strands are additionally linked by a segment of single stranded nucleotides so that the first and second strands are parts of a single oligonucleotide chain. Kmiec I further teaches a method for introducing specific 20 genetic alterations into a target gene. According to Kmiec I, the sequences of the RNA segments are selected to be homologous, *i.e.*, identical, to the sequence of a first and a second fragment of the target gene. The sequence of the interposed DNA segment is homologous with the sequence of the target gene between the first and second fragment except for a region of difference, termed the “heterologous region.” The heterologous 25 region can effect an insertion or deletion, or can contain one or more bases that are mismatched with the sequence of target gene so as to effect a substitution. According to Kmiec I, the sequence of the target gene is altered as directed by the heterologous region, such that the target gene becomes homologous with the sequence of the recombinagenic oligonucleobase. Kmiec I specifically teaches that ribose and 2'-O-methylribose, *i.e.*, 2'- 30 methoxyribose, containing nucleotides can be used in recombinagenic oligonucleobases and that naturally-occurring deoxyribose-containing nucleotides can be used as DNA-like nucleotides.

U.S. Patent No. 5,731,181 to Kmiec (Kmiec II) specifically disclose the use of recombinagenic oligonucleobases to effect genetic changes in plant cells and discloses 35 further examples of analogs and derivatives of RNA-like and DNA-like nucleotides that can be used to effect genetic changes in specific target genes. Other patents discussing the use

of recombinagenic oligonucleobases include: U.S. Patent Nos. 5,756,325; 5,871,984; 5,760,012; 5,888,983; 5,795,972; 5,780,296; 5,945,339; 6,004,804; and 6,010,907 and in International Patent No. PCT/US00/23457; and in International Patent Publication Nos. WO 98/49350; WO 99/07865; WO 99/58723; WO 99/58702; and WO 99/40789.

5 Recombinagenic oligonucleobases include mixed duplex oligonucleotides, non-nucleotide containing molecules taught in Kmiec II and other molecules taught in the above-noted patents and patent publications.

Citation or identification of any reference in Section 2, or any section of this application shall not be construed as an admission that such reference is available as prior art to the present invention.

3. SUMMARY OF THE INVENTION

The present invention is directed to a non-transgenic plant or plant cell having one or more mutations in the EPSPS gene, which plant has increased resistance or tolerance to a member of the phosphonomethylglycine family and which plant exhibits substantially normal growth or development of the plant, its organs, tissues or cells, as compared to the corresponding wild-type plant or cell. The present invention is also directed to a non-transgenic plant having a mutation in the EPSPS gene, which plant is resistant to or has an increased tolerance to a member of the phosphonomethylglycine family, *e.g.*, glyphosate, wherein the mutated EPSPS protein has substantially the same catalytic activity as compared to the wild-type EPSPS protein.

The present invention is also directed to a method for producing a non-transgenic plant having a mutated EPSPS gene that substantially maintains the catalytic activity of the wild-type protein irrespective of the presence or absence of a herbicide of the phosphonomethylglycine family. The method comprises introducing into a plant cell a recombinagenic oligonucleobase with a targeted mutation in the EPSPS gene and identifying a cell, seed, or plant having a mutated EPSPS gene.

Illustrative examples of a recombinagenic oligonucleobase is found in following patent publications, which are incorporated in their entirety by reference herein:

30 U.S. Patent Nos. 5,565,350; 5,756,325; 5,871,984; 5,760,012; 5,731,181; 5,888,983; 5,795,972; 5,780,296; 5,945,339; 6,004,804; and 6,010,907 and in International Patent No. PCT/US00/23457; and in International Patent Publication Nos. WO 98/49350; WO 99/07865; WO 99/58723; WO 99/58702; and WO 99/40789.

The plant can be of any species of dicotyledonous, monocotyledonous or 35 gymnospermous plant, including any woody plant species that grows as a tree or shrub, any herbaceous species, or any species that produces edible fruits, seeds or vegetables, or any

species that produces colorful or aromatic flowers. For example, the plant may be selected from a species of plant from the group consisting of canola, sunflower, tobacco, sugar beet, cotton, maize, wheat, barley, rice, sorghum, tomato, mango, peach, apple, pear, strawberry, banana, melon, potato, carrot, lettuce, onion, soya spp, sugar cane, pea, field beans, poplar, 5 grape, citrus, alfalfa, rye, oats, turf and forage grasses, flax, oilseed rape, cucumber, morning glory, balsam, pepper, eggplant, marigold, lotus, cabbage, daisy, carnation, tulip, iris, lily, and nut producing plants insofar as they are not already specifically mentioned.

The recombinogenic oligonucleobase can be introduced into a plant cell using any method commonly used in the art, including but not limited to, microcarriers 10 (biolistic delivery), microfibers, electroporation, microinjection.

The invention is also directed to the culture of cells mutated according to the methods of the present invention in order to obtain a plant that produces seeds, henceforth a “fertile plant”, and the production of seeds and additional plants from such a fertile plant.

The invention is further directed to a method of selectively controlling weeds 15 in a field, the field comprising plants with the disclosed EPSPS gene alterations and weeds, the method comprising application to the field of a herbicide to which the said plants have been rendered resistant.

The invention is also directed to novel mutations in the EPSPS gene that confer resistance or tolerance to a member of the phosphonomethylglycine family, *e.g.*, 20 glyphosate, to a plant or wherein the mutated EPSPS has substantially the same enzymatic activity as compared to wild-type EPSPS.

3.1 DEFINITIONS

The invention is to be understood in accordance with the following 25 definitions.

An oligonucleobase is a polymer of nucleobases, which polymer can hybridize by Watson-Crick base pairing to a DNA having the complementary sequence.

Nucleobases comprise a base, which is a purine, pyrimidine, or a derivative or analog thereof. Nucleobases include peptide nucleobases, the subunits of peptide nucleic acids, and morpholine nucleobases as well as nucleosides and nucleotides. Nucleosides are nucleobases that contain a pentosefuranosyl moiety, *e.g.*, an optionally substituted riboside or 2'-deoxyriboside. Nucleosides can be linked by one of several linkage moieties, which may or may not contain a phosphorus. Nucleosides that are linked by unsubstituted phosphodiester linkages are termed nucleotides.

35 An oligonucleobase chain has a single 5' and 3' terminus, which are the ultimate nucleobases of the polymer. A particular oligonucleobase chain can contain

nucleobases of all types. An oligonucleobase compound is a compound comprising one or more oligonucleobase chains that are complementary and hybridized by Watson-Crick base pairing. Nucleobases are either deoxyribo-type or ribo-type. Ribo-type nucleobases are pentosefuranosyl containing nucleobases wherein the 2' carbon is a methylene substituted 5 with a hydroxyl, alkyloxy or halogen. Deoxyribo-type nucleobases are nucleobases other than ribo-type nucleobases and include all nucleobases that do not contain a pentosefuranosyl moiety.

An oligonucleobase strand generically includes both oligonucleobase chains and segments or regions of oligonucleobase chains. An oligonucleobase strand has a 3' end 10 and a 5' end. When a oligonucleobase strand is coextensive with a chain, the 3' and 5' ends of the strand are also 3' and 5' termini of the chain.

According to the present invention, substantially normal growth of a plant, plant organ, plant tissue or plant cell is defined as a growth rate or rate of cell division of the plant, plant organ, plant tissue, or plant cell that is at least 35%, at least 50%, at least 60%, 15 or at least 75% of the growth rate or rate of cell division in a corresponding plant, plant organ, plant tissue or plant cell expressing the wild type EPSPS protein.

According to the present invention, substantially normal development of a plant, plant organ, plant tissue or plant cell is defined as the occurrence of one or more developmental events in the plant, plant organ, plant tissue or plant cell that are 20 substantially the same as those occurring in a corresponding plant, plant organ, plant tissue or plant cell expressing the wild type EPSPS protein.

According to the present invention plant organs include, but are not limited to, leaves, stems, roots, vegetative buds, floral buds, meristems, embryos, cotyledons, endosperm, sepals, petals, pistils, carpels, stamens, anthers, microspores, pollen, pollen 25 tubes, ovules, ovaries and fruits, or sections, slices or discs taken therefrom. Plant tissues include, but are not limited to, callus tissues, ground tissues, vascular tissues, storage tissues, meristematic tissues, leaf tissues, shoot tissues, root tissues, gall tissues, plant tumor tissues, and reproductive tissues. Plant cells include, but are not limited to, isolated cells with cell walls, variously sized aggregates thereof, and protoplasts.

30 Plants are substantially “tolerant” to glyphosate when they are subjected to it and provide a dose/response curve which is shifted to the right when compared with that provided by similarly subjected non-tolerant like plant. Such dose/response curves have “dose” plotted on the X-axis and “percentage kill”, “herbicidal effect”, etc., plotted on the y-axis. Tolerant plants will require more herbicide than non-tolerant like plants in order to 35 produce a given herbicidal effect. Plants which are substantially “resistant” to the glyphosate exhibit few, if any, necrotic, lytic, chlorotic or other lesions, when subjected to

glyphosate at concentrations and rates which are typically employed by the agrochemical community to kill weeds in the field. Plants which are resistant to a herbicide are also tolerant of the herbicide. The terms “resistant” and “tolerant” are to be construed as “tolerant and/or resistant” within the context of the present application.

5

4. BRIEF DESCRIPTION OF THE FIGURES

FIG. 1A is the DNA sequence of *Arabidopsis thaliana* EPSPS gene (SEQ ID NO:1). The bold underlined nucleotide residues are the targeted residues.

FIG. 1B is the amino acid sequence of *Arabidopsis thaliana* EPSPS protein (SEQ ID NO:2). The bold and underlined amino acid residues are the targeted residues.

FIG. 2 is a list of the *Arabidopsis thaliana* wild-type and mutant EPSPS nucleotide and amino acid sequences in the region of amino acid position 173 to 183; wild-type nucleotide sequence (SEQ ID NO:1) and wild-type amino acid sequence (SEQ ID NO:2), mutant A₁₇₇ nucleotide sequence (SEQ ID NO:3) and amino acid sequence (SEQ ID NO:4); mutant I₁₇₈ nucleotide sequence (SEQ ID NO:5) and amino acid sequence (SEQ ID NO:6); mutant A₁₇₇I₁₇₈ nucleotide sequence (SEQ ID NO:7) and amino acid sequence (SEQ ID NO:8); mutant I₁₇₈S₁₈₂ nucleotide sequence (SEQ ID NO:9) and amino acid sequence (SEQ ID NO:10); mutant A₁₇₇S₁₈₂ nucleotide sequence (SEQ ID NO:11) and amino acid sequence (SEQ ID NO:12); mutant A₁₇₇I₁₇₈S₁₈₂ nucleotide sequence (SEQ ID NO:13) and amino acid sequence (SEQ ID NO:14); mutant V₁₇₇S₁₈₂ nucleotide sequence (SEQ ID NO:15) and amino acid sequence (SEQ ID NO:16); mutant L₁₇₈S₁₈₂ nucleotide sequence (SEQ ID NO:17) and amino acid sequence (SEQ ID NO:18); mutant A₁₇₇V₁₇₈ nucleotide sequence (SEQ ID NO:19) and amino acid sequence (SEQ ID NO:20); mutant A₁₇₇L₁₈₂ nucleotide sequence (SEQ ID NO:21) and amino acid sequence (SEQ ID NO:22).

FIG. 3A-C is an alignment of the DNA of *Arabidopsis thaliana* EPSPS gene performed by DNAStar (LaserGene), (SEQ ID NO:1) with the nucleotide sequences of *Brassica napus* (SEQ ID NO:23); *Petunia hybrida* (SEQ ID NO:24); and *Zea mays* (SEQ ID NO:25) EPSPS gene. The sequences are aligned using J. Hein method with weighted residue weight table.

30

FIG. 4 is an alignment of the *Arabidopsis thaliana* EPSPS amino acid sequence (SEQ ID NO:2) with the *Brassica napus* (SEQ ID NO:26); *Petunia hybrida* (SEQ ID NO:27); and *Zea mays* (SEQ ID NO:28) EPSPS amino acid sequences. The sequences are aligned using J. Hein method with weighted residue weight table.

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FIG. 5 is a list of the mutagenesis primers used, with the targeted codons in bold characters (mutant primer A₁₇₇ (SEQ ID NO:29); mutant primer I₁₇₈ (SEQ ID NO:30);

mutant primer A₁₇₇I₁₇₈ (SEQ ID NO:31); mutant primer I₁₇₈S₁₈₂ (SEQ ID NO:32); mutant primer A₁₇₇S₁₈₂ (SEQ ID NO:34); mutant primer A₁₇₇I₁₇₈S₁₈₂ (SEQ ID NO:35); mutant primer V₁₇₇S₁₈₂ (SEQ ID NO:35); mutant primer L₁₇₈S₁₈₂ (SEQ ID NO:36); mutant primer A₁₇₇V₁₇₈ (SEQ ID NO:37); and mutant primer A₁₇₇L₁₈₂ (SEQ ID NO:38)).

5 FIG. 6 is the growth measured by optical density at 600 nm of *Arabidopsis* clones in the presence (+) and absence (-) of 17 mM glyphosate.

FIG. 7 (top panel) is a western blot showing the expression of His-tagged *Bacillus*, *Arabidopsis* wild type (WT) and mutant (AS) EPSPS proteins isolated from cell lysates (L) and eluates (E). Untransformed *Salmonella* as a negative control shows no 10 EPSPS expression. The bottom panel is a silver-stained duplicate gel.

5. DETAILED DESCRIPTION OF THE INVENTION

The present invention is directed to a non-transgenic plant or plant cell having a mutation in the EPSPS gene, which plant has increased resistance or tolerance to a 15 member of the phosphonomethylglycine family and which plant exhibits substantially normal growth or development of the plant, its organs, tissues or cells, as compared to the corresponding wild-type plant or cell. The present invention is also directed to a non-transgenic plant having a mutation in the EPSPS gene, which plant is resistant to or has an increased tolerance to a member of the phosphonomethylglycine family, *e.g.*, glyphosate, 20 wherein the mutated EPSPS protein has substantially the same catalytic activity as compared to the wild-type EPSPS protein.

The present invention is also directed to a method for producing a non-transgenic plant having a mutated EPSPS gene that substantially maintains the catalytic activity of the wild-type protein irrespective of the presence or absence of a herbicide of the 25 phosphonomethylglycine family. The method comprises introducing into a plant cell a recombinagenic oligonucleobase with a targeted mutation in the EPSPS gene and identifying a cell, seed, or plant having a mutated EPSPS gene.

Illustrative examples of a recombinagenic oligonucleobase is found in following patent publications, which are incorporated in their entirety by reference herein: 30 U.S. Patent Nos. 5,565,350; 5,756,325; 5,871,984; 5,760,012; 5,731,181; 5,888,983; 5,795,972; 5,780,296; 5,945,339; 6,004,804; and 6,010,907 and in International Patent No. PCT/US00/23457; and in International Patent Publication Nos. WO 98/49350; WO 99/07865; WO 99/58723; WO 99/58702; and WO 99/40789.

The plant can be of any species of dicotyledonous, monocotyledonous or 35 gymnospermous plant, including any woody plant species that grows as a tree or shrub, any herbaceous species, or any species that produces edible fruits, seeds or vegetables, or any

species that produces colorful or aromatic flowers. For example, the plant may be selected from a species of plant from the group consisting of canola, sunflower, tobacco, sugar beet, cotton, maize, wheat, barley, rice, sorghum, tomato, mango, peach, apple, pear, strawberry, banana, melon, potato, carrot, lettuce, onion, soya spp, sugar cane, pea, field beans, poplar, 5 grape, citrus, alfalfa, rye, oats, turf and forage grasses, flax, oilseed rape, cucumber, morning glory, balsam, pepper, eggplant, marigold, lotus, cabbage, daisy, carnation, tulip, iris, lily, and nut producing plants insofar as they are not already specifically mentioned.

The recombinagenic oligonucleobase can be introduced into a plant cell using any method commonly used in the art, including but not limited to, microcarriers 10 (biolistic delivery), microfibers, electroporation, microinjection.

The invention is also directed to the culture of cells mutated according to the methods of the present invention in order to obtain a plant that produces seeds, henceforth a “fertile plant”, and the production of seeds and additional plants from such a fertile plant.

The invention is further directed to a method of selectively controlling weeds 15 in a field, the field comprising plants with the disclosed EPSPS gene alterations and weeds, the method comprising application to the field of a herbicide to which the said plants have been rendered resistant.

The invention is also directed to novel mutations in the EPSPS gene that confer resistance or tolerance to a member of the phosphonomethylglycine family, *e.g.*, 20 glyphosate, to a plant or wherein the mutated EPSPS has substantially the same enzymatic activity as compared to wild-type EPSPS.

5.1 RECOMBINAGENIC OLIGONUCLEOBASES

The invention can be practiced with recombinagenic oligonucleobases 25 having the conformations and chemistries described in United States patent No. 5,565,350 to Kmiec (Kmiec I) and U.S. patent No. 5,731,181 (Kmiec II) gene, which are hereby incorporated by reference. Kmiec I teaches a method for introducing specific genetic alterations into a target gene. The recombinagenic oligonucleobases in Kmiec I and/or Kmiec II contain two complementary strands, one of which contains at least one segment of 30 RNA-type nucleotides (an “RNA segment”) that are base paired to DNA-type nucleotides of the other strand.

Kmiec II discloses that purine and pyrimidine base-containing non-nucleotides can be substituted for nucleotides. U.S. Patent Nos. 5,756,325; 5,871,984; 5,760,012; 5,888,983; 5,795,972; 5,780,296; 5,945,339; 6,004,804; and 6,010,907 and in 35 International Patent No. PCT/US00/23457; and in International Patent Publication Nos. WO 98/49350; WO 99/07865; WO 99/58723; WO 99/58702; and WO 99/40789, which are each

hereby incorporated in their entirety, disclose additional recombinagenic molecules that can be used for the present invention. The term “recombinagenic oligonucleobase” is used herein to denote the molecules that can be used in the methods of the present invention and include mixed duplex oligonucleotides, non-nucleotide containing molecules taught in 5 Kmiec II, single stranded oligodeoxynucleotides and other recombinagenic molecules taught in the above noted patents and patent publications.

In one embodiment, the recombinagenic oligonucleobase is a mixed duplex oligonucleotide in which the RNA-type nucleotides of the mixed duplex oligonucleotide are made RNase resistant by replacing the 2'-hydroxyl with a fluoro, chloro or bromo 10 functionality or by placing a substituent on the 2'-O. Suitable substituents include the substituents taught by the Kmiec II. Alternative substituents include the substituents taught by U.S. Patent No. 5,334,711 (Sproat) and the substituents taught by patent publications EP 629 387 and EP 679 657 (collectively, the Martin Applications), which are hereby 15 incorporated by reference. As used herein, a 2'-fluoro, chloro or bromo derivative of a ribonucleotide or a ribonucleotide having a 2'-OH substituted with a substituent described in the Martin Applications or Sproat is termed a “2'-Substituted Ribonucleotide.” As used herein the term “RNA-type nucleotide” means a 2'-hydroxyl or 2'-Substituted Nucleotide that is linked to other nucleotides of a mixed duplex oligonucleotide by an unsubstituted phosphodiester linkage or any of the non-natural linkages taught by Kmiec I or Kmiec II. 20 As used herein the term “deoxyribo-type nucleotide” means a nucleotide having a 2'-H, which can be linked to other nucleotides of a MDON by an unsubstituted phosphodiester linkage or any of the non-natural linkages taught by Kmiec I or Kmiec II.

In a particular embodiment of the present invention, the recombinagenic oligonucleobase is a mixed duplex oligonucleotide that is linked solely by unsubstituted 25 phosphodiester bonds. In alternative embodiments, the linkage is by substituted phosphodiesters, phosphodiester derivatives and non-phosphorus-based linkages as taught by Kmiec II. In yet another embodiment, each RNA-type nucleotide in the mixed duplex oligonucleotide is a 2'-Substituted Nucleotide. Particular preferred embodiments of 2'-Substituted Ribonucleotides are 2'-fluoro, 2'-methoxy, 2'-propyloxy, 2'-allyloxy, 2'- 30 hydroxylethoxy, 2'-methoxyethoxy, 2'-fluoropropoxy and 2'-trifluoropropoxy substituted ribonucleotides. More preferred embodiments of 2'-Substituted Ribonucleotides are 2'-fluoro, 2'-methoxy, 2'-methoxyethoxy, and 2'-allyloxy substituted nucleotides. In another embodiment the mixed duplex oligonucleotide is linked by unsubstituted phosphodiester bonds.

35 Although mixed duplex oligonucleotide having only a single type of 2'-substituted RNA-type nucleotide are more conveniently synthesized, the methods of the

invention can be practiced with mixed duplex oligonucleotides having two or more types of RNA-type nucleotides. The function of an RNA segment may not be affected by an interruption caused by the introduction of a deoxynucleotide between two RNA-type trinucleotides, accordingly, the term RNA segment encompasses such an "interrupted RNA 5 segment." An uninterrupted RNA segment is termed a contiguous RNA segment. In an alternative embodiment an RNA segment can contain alternating RNase-resistant and unsubstituted 2'-OH nucleotides. The mixed duplex oligonucleotides preferably have fewer than 100 nucleotides and more preferably fewer than 85 nucleotides, but more than 50 nucleotides. The first and second strands are Watson-Crick base paired. In one 10 embodiment the strands of the mixed duplex oligonucleotide are covalently bonded by a linker, such as a single stranded hexa, penta or tetranucleotide so that the first and second strands are segments of a single oligonucleotide chain having a single 3' and a single 5' end. The 3' and 5' ends can be protected by the addition of a "hairpin cap" whereby the 3' and 5' terminal nucleotides are Watson-Crick paired to adjacent nucleotides. A second hairpin cap 15 can, additionally, be placed at the junction between the first and second strands distant from the 3' and 5' ends, so that the Watson-Crick pairing between the first and second strands is stabilized.

The first and second strands contain two regions that are homologous with two fragments of the target EPSPS gene, *i.e.*, have the same sequence as the target gene. A 20 homologous region contains the nucleotides of an RNA segment and may contain one or more DNA-type nucleotides of connecting DNA segment and may also contain DNA-type nucleotides that are not within the intervening DNA segment. The two regions of homology are separated by, and each is adjacent to, a region having a sequence that differs from the sequence of the target gene, termed a "heterologous region." The heterologous 25 region can contain one, two or three mismatched nucleotides. The mismatched nucleotides can be contiguous or alternatively can be separated by one or two nucleotides that are homologous with the target gene. Alternatively, the heterologous region can also contain an insertion or one, two, three or of five or fewer nucleotides. Alternatively, the sequence of the mixed duplex oligonucleotide may differ from the sequence of the target gene only by 30 the deletion of one, two, three, or five or fewer nucleotides from the mixed duplex oligonucleotide. The length and position of the heterologous region is, in this case, deemed to be the length of the deletion, even though no nucleotides of the mixed duplex oligonucleotide are within the heterologous region. The distance between the fragments of the target gene that are complementary to the two homologous regions is identically the 35 length of the heterologous region when a substitution or substitutions is intended. When the heterologous region contains an insertion, the homologous regions are thereby separated in

the mixed duplex oligonucleotide farther than their complementary homologous fragments are in the gene, and the converse is applicable when the heterologous region encodes a deletion.

The RNA segments of the mixed duplex oligonucleotides are each a part of a homologous region, *i.e.*, a region that is identical in sequence to a fragment of the target gene, which segments together preferably contain at least 13 RNA-type nucleotides and preferably from 16 to 25 RNA-type nucleotides or yet more preferably 18-22 RNA-type nucleotides or most preferably 20 nucleotides. In one embodiment, RNA segments of the homology regions are separated by and adjacent to, *i.e.*, “connected by” an intervening DNA segment. In one embodiment, each nucleotide of the heterologous region is a nucleotide of the intervening DNA segment. An intervening DNA segment that contains the heterologous region of a mixed duplex oligonucleotide is termed a “mutator segment.”

The change to be introduced into the target EPSPS gene is encoded by the heterologous region. The change to be introduced into the EPSPS gene may be a change in one or more bases of the EPSPS gene sequence or the addition or deletion of one or more bases.

In another embodiment of the present invention, the recombinagenic oligonucleobase is a single stranded oligodeoxynucleotide mutational vector or SSOMV, which is disclosed in International Patent Application PCT/US00/23457, which is incorporated by reference in its entirety. The sequence of the SSOMV is based on the same principles as the mutational vectors described in U.S. Patent Nos. 5,756,325; 5,871,984; 5,760,012; 5,888,983; 5,795,972; 5,780,296; 5,945,339; 6,004,804; and 6,010,907 and in International Publication Nos. WO 98/49350; WO 99/07865; WO 99/58723; WO 99/58702; and WO 99/40789. The sequence of the SSOMV contains two regions that are homologous with the target sequence separated by a region that contains the desired genetic alteration termed the mutator region. The mutator region can have a sequence that is the same length as the sequence that separates the homologous regions in the target sequence, but having a different sequence. Such a mutator region can cause a substitution. Alternatively, the homologous regions in the SSOMV can be contiguous to each other, while the regions in the target gene having the same sequence are separated by one, two or more nucleotides. Such a SSOMV causes a deletion from the target gene of the nucleotides that are absent from the SSOMV. Lastly, the sequence of the target gene that is identical to the homologous regions may be adjacent in the target gene but separated by one two or more nucleotides in the sequence of the SSOMV. Such an SSOMV causes an insertion in the sequence of target gene.

The nucleotides of the SSOMV are deoxyribonucleotides that are linked by unmodified phosphodiester bonds except that the 3' terminal and/or 5' terminal internucleotide linkage or alternatively the two 3' terminal and/or 5' terminal internucleotide linkages can be a phosphorothioate or phosphoamidate. As used herein an internucleotide linkage is the linkage between nucleotides of the SSOMV and does not include the linkage between the 3' end nucleotide or 5' end nucleotide and a blocking substituent, see *supra*. In a specific embodiment the length of the SSOMV is between 21 and 55 deoxynucleotides and the lengths of the homology regions are, accordingly, a total length of at least 20 deoxynucleotides and at least two homology regions should each have lengths of at least 8 deoxynucleotides.

The SSOMV can be designed to be complementary to either the coding or the non-coding strand of the target gene. When the desired mutation is a substitution of a single base, it is preferred that both the mutator nucleotide be a pyrimidine. To the extent that is consistent with achieving the desired functional result it is preferred that both the mutator nucleotide and the targeted nucleotide in the complementary strand be pyrimidines. Particularly preferred are SSOMV that encode transversion mutations, *i.e.*, a C or T mutator nucleotide is mismatched, respectively, with a C or T nucleotide in the complementary strand.

In addition to the oligodeoxynucleotide the SSOMV can contain a 5' blocking substituent that is attached to the 5' terminal carbons through a linker. The chemistry of the linker is not critical other than its length, which should preferably be at least 6 atoms long and that the linker should be flexible. A variety of non-toxic substituents such as biotin, cholesterol or other steroids or a non-intercalating cationic fluorescent dye can be used. Particularly preferred as reagents to make SSOMV are the reagents sold as Cy3™ and Cy5™ by Glen Research, Sterling VA, which are blocked phosphoroamidites that upon incorporation into an oligonucleotide yield 3,3,3',3'-tetramethyl N,N'-isopropyl substituted indomonocarbocyanine and indodicarbocyanine dyes, respectively. Cy3 is the most preferred. When the indocarbocyanine is N-oxyalkyl substituted it can be conveniently linked to the 5' terminal of the oligodeoxynucleotide through a phosphodiester with a 5' terminal phosphate. The chemistry of the dye linker between the dye and the oligodeoxynucleotide is not critical and is chosen for synthetic convenience. When the commercially available Cy3 phosphoramidite is used as directed the resulting 5' modification consists of a blocking substituent and linker together which are a N-hydroxypropyl, N'-phosphatidylpropyl 3,3,3',3'-tetramethyl indomonocarbocyanine.

In the preferred embodiment the indocarbocyanine dye is tetra substituted at the 3 and 3' positions of the indole rings. Without limitation as to theory these substitutions

prevent the dye from being an intercalating dye. The identity of the substituents at these positions are not critical. The SSOMV can in addition have a 3' blocking substituent. Again the chemistry of the 3' blocking substituent is not critical.

5

5.2 THE LOCATION AND TYPE OF MUTATION INTRODUCED INTO THE EPSPS GENE

In one embodiment of the present invention, the *Arabidopsis thaliana* EPSPS gene (see Figure 1A) and corresponding EPSPS enzyme (see Figure 1B) comprises a mutation at one or more amino acid residues selected from the group consisting of Leu₁₇₃, Gly₁₇₇, Thr₁₇₈, Ala₁₇₉, Met₁₈₀, Arg₁₈₁, Pro₁₈₂, Ser₉₈, Ser₂₅₅ and Leu₁₉₈, or at an analogous 10 position in an EPSPS paralog, and the mutation results in one or more of the following amino acid substitutions in the EPSPS enzyme in comparison with the wild-type sequence:

- 15 (i) Leu₁₇₃ - Phe
- (ii) Gly₁₇₇ - Ala or Ile
- (iii) Thr₁₇₈ - Ile or Val or Leu
- (iv) Ala₁₇₉ - Gly
- (v) Met₁₈₀ - Cys
- (vi) Arg₁₈₁ - Leu or Ser
- (vii) Pro₁₈₂ - Leu or Ser
- 20 (viii) Ser₉₈ -Asp
- (ix) Ser₂₅₅ -Ala
- (x) Leu₁₉₈ -Lys.

In another embodiment of the present invention, within the EPSPS gene product, the amino acid residue to be changed is Leu within the contiguous sequence Leu-25 Tyr-Leu-Gly-Asn (SEQ ID NO:29) and is changed to Phe; or the amino acid residue to be changed is Gly within the contiguous sequence Asn-Ala-Gly-Thr-Ala (SEQ ID NO:30) and is changed to Ala or Ile; or the amino acid to be changed is Thr within the contiguous sequence Ala-Gly-Thr-Ala-Met (SEQ ID NO:31) and is changed to Ile, Val or Leu; or the amino acid to be changed is Ala within the contiguous sequence Gly-Thr-Ala-Met-Arg 30 (SEQ ID NO:32) and is changed to Gly; or the amino acid to be changed is Met within the contiguous sequence Thr-Ala-Met-Arg-Pro (SEQ ID NO:33) and is changed to Cys; or the amino acid to be changed is Arg within the contiguous sequence Ala-Met-Arg-Pro-Leu (SEQ ID NO:34) and is changed to Leu or Ser; or the amino acid to be changed is Pro within the contiguous sequence Met-Arg-Pro-Leu-Thr (SEQ ID NO:35) and is changed to 35 Leu or Ser; or the amino acid to be changed is Ser within a contiguous Pro-Gly-Ser-Lys-Ser (SEQ ID NO:36) and is changed to Asp; or the amino acid to be changed is Ser within the

contiguous sequence Ile-Ser-Ser-Gln-Tyr (SEQ ID NO:37) and is changed to Ala; or the amino acid to be changed is Leu within the contiguous sequence Tyr-Val-Leu-Asp-Gly (SEQ ID NO:38) and is changed to Lys. In other embodiments, one or more of the foregoing changes can be made in the EPSPS amino acid sequence.

5 Alternatively, and/or additionally, the mutation may result in the replacement of any amino acid at positions corresponding to 256, 284-288 and 353-356 with respect to the EPSPS protein depicted in Figure 1B (SEQ ID NO. 2).

In specific embodiments of the present invention, the EPSPS gene is mutated at amino acid position 177 in which Gly is replaced by Ala. Another specific embodiment 10 is the substitution of Thr at amino acid position 178 by Ile. A further specific embodiment comprises a mutation at amino acid position 177 in which Gly is replaced by Ala, plus the additional substitution of Thr at amino acid position 178 by Ile. Other specific 15 embodiments of the present invention are directed to mutations at amino acid position 178, in which Thr is replaced by Ile, plus the additional mutation at position 182, in which Pro is replaced by Ser. Other embodiments include the substitution of Gly at amino acid position 177 by Ala, plus the additional mutation at amino acid position 182, in which Pro is substituted by Ser. Other mutated EPSPS sequences comprise the substitution of Gly at position 177 by Ala, plus the substitution at position 178, in which Thr is replaced by Ile, plus the additional substitution of Pro at amino acid position 182 by Ser. Another 20 embodiment is the substitution of Thr at amino acid position 178 by Val and the additional mutation at amino acid position 182, in which Pro is replaced by Ser. A further specific embodiment includes the substitution of Thr at position 178 by Leu, plus the mutation at amino acid position 182, in which Pro is replaced by Ser. A further embodiment includes, the substitution at amino acid position 177 in which Gly is replaced by Ala, plus the 25 substitution of Thr at position 178 by Val. The invention also embodies the substitution at amino acid position 177 in which Gly is replaced by Ala, plus the replacement of Thr at amino acid position 178 by Leu (see Figure 2).

The foregoing mutations in the EPSPS gene were described using the *Arabidopsis thaliana* EPSPS gene (SEQ ID NO:1) and protein (SEQ ID NO:2). The 30 present invention also encompasses mutant EPSPS genes of other species (paralogs). However, due to variations in the EPSPS genes of different species, the number of the amino acid residue to be changed in one species may be different in another species. Nevertheless, the analogous position is readily identified by one of skill in the art by sequence homology. For example, Figure 3A-C shows the aligned nucleotide sequences 35 and Figure 4 shows the aligned amino acid sequences of four paralogs of the EPSPS gene, *Arabidopsis thaliana*, *Zea mays*, *Petunia hybrida*, and *Brassica napus*. Thus, the analogous

positions in *Zea mays* are Leu₉₇, Gly₁₀₁, Thr₁₀₂, Ala₁₀₃, Met₁₀₄, Arg₁₀₅, Pro₁₀₆, Ser₂₃, Ser₁₇₉ and Leu₁₂₂. Thus, the *Zea mays* EPSPS amino acid sequence is mutated at one or more of the following amino acid positions and results in one or more of the following substitutions:

- (i) Leu₉₇ - Phe
- 5 (ii) Gly₁₀₁ - Ala or Ile
- (iii) Thr₁₀₂ - Ile or Val or Leu
- (iv) Ala₁₀₃ - Gly
- (v) Met₁₀₄ - Cys
- (vi) Arg₁₀₅ - Leu or Ser
- 10 (vii) Pro₁₀₆ - Leu or Ser
- (viii) Ser₂₃ - Asp
- (ix) Ser₁₇₉ - Ala
- (x) Leu₁₂₂ - Lys.

In another embodiment of the present invention, within the *Zea mays* EPSPS gene product the amino acid residue to be changed is Leu within the contiguous sequence Leu-Phe-Leu-Gly-Asn (SEQ ID NO:39) and is changed to Phe; or the amino acid residue to be changed is Gly within the contiguous sequence Asn-Ala-Gly-Thr-Ala (SEQ ID NO:30) and is changed to Ala or Ile; or the amino acid to be changed is Thr within the contiguous sequence Ala-Gly-Thr-Ala-Met (SEQ ID NO:31) and is changed to Ile, Val or Leu; or the amino acid to be changed is Ala within the contiguous sequence Gly-Thr-Ala-Met-Arg (SEQ ID NO:32) and is changed to Gly; or the amino acid to be changed is Met within the contiguous sequence Thr-Ala-Met-Arg-Pro (SEQ ID NO:33) and is changed to Cys; or the amino acid to be changed is Arg within the contiguous sequence Ala-Met-Arg-Pro-Leu (SEQ ID NO:34) and is changed to Leu or Ser; or the amino acid to be changed is Pro within the contiguous sequence Met-Arg-Pro-Leu-Thr (SEQ ID NO:35) and is changed to Leu or Ser; or the amino acid to be changed is Ser within a contiguous Pro-Gly-Ser-Lys-Ser (SEQ ID NO:36) and is changed to Asp; or the amino acid to be changed is Ser within the contiguous sequence Ile-Ser-Ser-Gln-Tyr (SEQ ID NO:37) and is changed to Ala; or the amino acid to be changed is Leu within the contiguous sequence Tyr-Val-Leu-Asp-Gly (SEQ ID NO:38) and is changed to Lys. In other embodiments, one or more of the foregoing changes can be made in the EPSPS amino acid sequence.

In *Brassica napus*, the analogous amino acid positions are Leu₁₆₉, Gly₁₇₃, Thr₁₇₄, Ala₁₇₅, Met₁₇₆, Arg₁₇₇, Pro₁₇₈, Ser₉₄, Ser₂₅₁ and Leu₁₉₄. Thus, the *Brassica napus* EPSPS amino acid sequence is mutated at one or more of the following amino acid positions and results in one or more of the following substitutions:

- (i) Leu₁₆₉ - Phe

5 (ii) Gly₁₇₃ - Ala or Ile
 (iii) Thr₁₇₄ - Ile or Val or Leu
 (iv) Ala₁₇₅ - Gly
 (v) Met₁₇₆ - Cys
 (vi) Arg₁₇₇ - Leu or Ser
 (vii) Pro₁₇₈ - Leu or Ser
 (viii) Ser₉₄ -Asp
 (ix) Ser₂₅₁ -Ala
 (x) Leu₁₉₄ -Lys

10 In another embodiment of the present invention, within the *Brassica napus* EPSPS gene product the amino acid residue to be changed is Leu within the contiguous sequence Leu-Tyr-Leu-Gly-Asn (SEQ ID NO:29) and is changed to Phe; or the amino acid residue to be changed is Gly within the contiguous sequence Asn-Ala-Gly-Thr-Ala (SEQ ID NO:30) and is changed to Ala or Ile; or the amino acid to be changed is Thr within the 15 contiguous sequence Ala-Gly-Thr-Ala-Met (SEQ ID NO:31) and is changed to Ile, Val or Leu; or the amino acid to be changed is Ala within the contiguous sequence Gly-Thr-Ala-Met-Arg (SEQ ID NO:32) and is changed to Gly; or the amino acid to be changed is Met within the contiguous sequence Thr-Ala-Met-Arg-Pro (SEQ ID NO:33) and is changed to Cys; or the amino acid to be changed is Arg within the contiguous sequence Ala-Met-Arg- 20 Pro-Leu (SEQ ID NO:34) and is changed to Leu or Ser; or the amino acid to be changed is Pro within the contiguous sequence Met-Arg-Pro-Leu-Thr (SEQ ID NO:35) and is changed to Leu or Ser; or the amino acid to be changed is Ser within a contiguous Pro-Gly-Ser-Lys-Ser (SEQ ID NO:36) and is changed to Asp; or the amino acid to be changed is Ser within the contiguous sequence Ile-Ser-Ser-Gln-Tyr (SEQ ID NO:37) and is changed to Ala; or the 25 amino acid to be changed is Leu within the contiguous sequence Tyr-Val-Leu-Asp-Gly (SEQ ID NO:38) and is changed to Lys. In other embodiments, one or more of the foregoing changes can be made in the EPSPS amino acid sequence.

In *Petunia hybrida* the analogous positions are Leu₁₆₉, Gly₁₇₃, Thr₁₇₄, Ala₁₇₅, Met₁₇₆, Arg₁₇₇, Pro₁₇₈, Ser₉₄, Ser₂₅₁ and Leu₁₉₄. Thus, the *Petunia hybrida* EPSPS amino acid sequence is mutated at one or more of the following amino acid positions and results in one or more of the following substitutions:

35 (i) Leu₁₆₉ - Phe
 (ii) Gly₁₇₃ - Ala or Ile
 (iii) Thr₁₇₄ - Ile or Val or Leu
 (iv) Ala₁₇₅ - Gly
 (v) Met₁₇₆ - Cys

- (vi) Arg₁₇₇ - Leu or Ser
- (vii) Pro₁₇₈ - Leu or Ser
- (viii) Ser₉₄ -Asp
- (ix) Ser₂₅₁ -Ala
- 5 (x) Leu₁₉₄ -Lys

In another embodiment of the present invention, within the *Petunia hybrida* EPSPS gene product the amino acid residue to be changed is Leu within the contiguous sequence Leu-Phe-Leu-Gly-Asn (SEQ ID NO:39) and is changed to Phe; or the amino acid residue to be changed is Gly within the contiguous sequence Asn-Ala-Gly-Thr-Ala (SEQ ID NO:30) and is changed to Ala or Ile; or the amino acid to be changed is Thr within the contiguous sequence Ala-Gly-Thr-Ala-Met (SEQ ID NO:31) and is changed to Ile, Val or Leu; or the amino acid to be changed is Ala within the contiguous sequence Gly-Thr-Ala-Met-Arg (SEQ ID NO:32) and is changed to Gly; or the amino acid to be changed is Met within the contiguous sequence Thr-Ala-Met-Arg-Pro (SEQ ID NO:33) and is changed to 10 Cys; or the amino acid to be changed is Arg within the contiguous sequence Ala-Met-Arg-Pro-Leu (SEQ ID NO:34) and is changed to Leu or Ser; or the amino acid to be changed is Pro within the contiguous sequence Met-Arg-Pro-Leu-Thr (SEQ ID NO:35) and is changed to Leu or Ser; or the amino acid to be changed is Ser within a contiguous Pro-Gly-Ser-Lys-Ser (SEQ ID NO:36) and is changed to Asp; or the amino acid to be changed is Ser within 15 the contiguous sequence Ile-Ser-Ser-Gln-Tyr (SEQ ID NO:37) and is changed to Ala; or the amino acid to be changed is Leu within the contiguous sequence Tyr-Val-Leu-Asp-Gly (SEQ ID NO:38) and is changed to Lys. In other embodiments, one or more of the foregoing changes can be made in the EPSPS amino acid sequence.

25 5.3 THE DELIVERY OF RECOMBINAGENIC OLIGONUCLEOBASES INTO PLANT CELLS

Any commonly known method can be used in the methods of the present invention to transform a plant cell with a recombinagenic oligonucleobases. Illustrative methods are listed below.

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5.3.1 MICROCARRIERS AND MICROFIBERS

The use of metallic microcarriers (microspheres) for introducing large fragments of DNA into plant cells having cellulose cell walls by projectile penetration is well known to those skilled in the relevant art (henceforth biolistic delivery). United States 35 Patent Nos. 4,945,050; 5,100,792 and 5,204,253 describe general techniques for selecting microcarriers and devices for projecting them.

Specific conditions for using microcarriers in the methods of the present invention are described in International Publication WO 99/07865. In an illustrative technique, ice cold microcarriers (60 mg/ml), mixed duplex oligonucleotide (60 mg/ml) 2.5 M CaCl₂ and 0.1 M spermidine are added in that order; the mixture gently agitated, *e.g.*, by 5 vortexing, for 10 minutes and let stand at room temperature for 10 minutes, whereupon the microcarriers are diluted in 5 volumes of ethanol, centrifuged and resuspended in 100% ethanol. Good results can be obtained with a concentration in the adhering solution of 8-10 μ g/ μ l microcarriers, 14-17 μ g/ml mixed duplex oligonucleotide, 1.1-1.4 M CaCl₂ and 18-10 22 mM spermidine. Optimal results were observed under the conditions of 8 μ g/ μ l microcarriers, 16.5 μ g/ml mixed duplex oligonucleotide, 1.3 M CaCl₂ and 21 mM spermidine.

Recombinagenic oligonucleobases can also be introduced into plant cells for the practice of the present invention using microfibers to penetrate the cell wall and cell membrane. U.S. Patent No. 5,302,523 to Coffee et al. describes the use of 30 x 0.5 μ m and 15 10 x 0.3 μ m silicon carbide fibers to facilitate transformation of suspension maize cultures of Black Mexican Sweet. Any mechanical technique that can be used to introduce DNA for transformation of a plant cell using microfibers can be used to deliver recombinagenic oligonucleobases for transmutation.

An illustrative technique for microfiber delivery of a recombinagenic oligonucleobase is as follows: Sterile microfibers (2 μ g) are suspended in 150 μ l of plant culture medium containing about 10 μ g of a mixed duplex oligonucleotide. A suspension culture is allowed to settle and equal volumes of packed cells and the sterile fiber/nucleotide suspension are vortexed for 10 minutes and plated. Selective media are applied immediately or with a delay of up to about 120 hours as is appropriate for the particular 25 trait.

5.3.2 PROTOPLAST ELECTROPORATION

In an alternative embodiment, the recombinagenic oligonucleobases can be delivered to the plant cell by electroporation of a protoplast derived from a plant part. The 30 protoplasts are formed by enzymatic treatment of a plant part, particularly a leaf, according to techniques well known to those skilled in the art. *See, e.g.*, Gallois et al., 1996, in Methods in Molecular Biology 55:89-107, Humana Press, Totowa, NJ; Kipp et al., 1999, in Methods in Molecular Biology 133:213-221, Humana Press, Totowa, NJ. The protoplasts need not be cultured in growth media prior to electroporation. Illustrative conditions for 35 electroporation are 3 x 10⁵ protoplasts in a total volume of 0.3 ml with a concentration of recombinagenic oligonucleobase of between 0.6 - 4 μ g/mL.

5.3.3 WHISKERS AND MICROINJECTION

In yet another alternative embodiment, the recombinagenic oligonucleobase can be delivered to the plant cell by whiskers or microinjection of the plant cell. The so called whiskers technique is performed essentially as described in Frame et al., 1994, Plant 5 J. 6:941-948. The recombinagenic oligonucleobase is added to the whiskers and used to transform the plant cells. The recombinagenic oligonucleobase may be co-incubated with plasmids comprising sequences encoding proteins capable of forming recombinase complexes in plant cells such that recombination is catalyzed between the oligonucleotide and the target sequence in the EPSPS gene.

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5.4 SELECTION OF GLYPHOSATE RESISTANT PLANTS

Plants or plant cells can be tested for resistance or tolerance to a herbicide using commonly known methods in the art, e.g., by growing the plant or plant cell in the presence of a herbicide and measuring the rate of growth as compared to the growth rate in 15 the absence of the herbicide.

6. EXAMPLE

The following experiments demonstrate the production of mutant *Arabidopsis thaliana* EPSPS genes which are resistant to the herbicide glyphosate and 20 which allows the plant cells to maintain a growth rate

6.1 MATERIAL AND METHODS

6.1.1 ISOLATION OF ARABIDOPSIS THALIANA EPSPS cDNA

A 1.3 kb DNA fragment was amplified by PCR from an *Arabidopsis* cDNA 25 library using the primers AtEXPEXPM1 and AtEXPEXP2CM-2. The two primers were designed to amplify the cDNA from the mature peptide to the termination codon. The 5' primer AtEXPEXPM1 contains an XbaI site (underlined) and the 3' primer AtEXPEXP2CM-2 contains a BglII site (underlined), sites which will be of use for cloning of the fragment into the expression vector.

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AtEXPEXPM1

5'-GCTCTAGAGAAAGCGTCGGAGATTGTACTT-3' (SEQ ID NO:40)

AtEXPEXP2CM-2

35 5'-GCAGATCTGAGCTCTAGTGCTTGTGATTCTTCAAGTAC-3' (SEQ ID NO:41)

The PCR band was excised from the agarose gel and purified (GeneClean, Biol). Its sequence was then confirmed as the mature peptide sequence of *Arabidopsis thaliana* EPSPS gene.

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6.1.2 PREPARATION OF THE EXPRESSION VECTOR

The EPSPS coding region of the *AroE* *Bacillus subtilis* gene was obtained by PCR using the following primers:

BsAroE5'Xba

5'-GCGTCTAGAAAAACGAGATAAGGTGCAG-3' (SEQ ID NO:42) and

10

BsAroE3'BamHI

5'-GCGGATCCTCAGGATTTTCGAAAGCTTATTAAATG-3' (SEQ ID NO:43).

15

The PCR fragment, lacking an initiation codon (ATG), was cloned in-frame to the pACLacIMH6RecA vector by replacing the ORF of *RecA* by digesting with *Xba*I and *Bam*HI. PACLacIMH6RecA contained the LacI region of Pet21 at positions 1440 to 3176, the MH6 RecA at positions 3809 to 5188, chloramphenicol resistance gene at positions 5445-218 (5446 to 5885 and 1 to 218), and the p15A origin of replication at positions 581 to 1424. The coding region of *RecA* gene was cloned from *E.coli* in-frame with the start codon and 6 histidine linker (MH6) behind the LacZ promoter of pUC19.

6.1.3 CLONING OF THE ARABIDOPSIS EPSPS GENE INTO BACTERIAL EXPRESSION VECTOR

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The *Arabidopsis* 1.3 kb PCR fragment was digested with *Xba*I and *Bam*HI (compatible with *Bgl*II) and cloned into the plasmid pACYCLacIMH6EPSPS, in place of the *Bacillus* gene.

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The clones obtained (selected on chloramphenicol) were then sequenced and confirmed positive. One of the confirmed clones (pAtEPS-12) was selected and the junctions between the cDNA and the cloning plasmid were also confirmed to be identical to the expected sequences.

6.1.4 NOVEL POINT MUTATIONS IN THE EPSPS GENE

Ten different mutants of the *Arabidopsis thaliana* EPSPS gene were designed, (see Figure 2). For the mutagenesis experiments, PCR primers were designed with one, two or three mutations. The PCR reactions were performed using a regular

flanking primer (5'ATEPS-198: 5'- GAAAGCGTCGGAGATTGTAC-3' (SEQ ID NO:44)) and one of the mutation-carrying primers (see Figure 5).

The 353bp PCR fragments obtained were purified (Qiagen PCR Purification kit) and their sequence confirmed. The fragments were then digested with PstI (underlined 5 in the primer sequences) and BamHI and ligated to the pAtEPS-12 vector, which had itself been previously digested with PstI and BamHI.JM109 (Promega) competent cells were used for the transformation and plated onto chloramphenicol-containing LB plates. Clones from each mutagenesis experiment were then isolated and their sequence confirmed.

10 6.1.5 GLYPHOSATE RESISTANCE ASSAYS

Electrocompetent cells of SA4247, a LacZ - *Salmonella typhi* strain, were prepared according to well known procedures (see Current Protocols in Molecular Biology, (Wiley and Sons, Inc.)). 30 μ l of SA4247 competent cells were electroporated with 20 ng of each plasmid DNA encoding *Arabidopsis* wild-type and mutant EPSPS proteins, *Bacillus* 15 wild-type EPSPS, along with a mock transfection as a control. The settings for electroporation were 25 μ F, 2.5KV and 200 ohms. After electroporation, the cells were transferred into 15 mls culture tube and supplemented with 970 μ l of SOC medium. The cultures were incubated for 1 $\frac{1}{2}$ hours at 37°C at 225 rpm. 50 μ l of each culture were plated onto LB plates containing 17 μ g/ml chloramphenicol (in duplicates) and incubated 20 overnight at 37°C. On the following day, 5 colonies of each plate were picked and transferred onto M9 plates and incubated overnight at 37°C.

Colonies from the overnight incubation on solid M9 were inoculated into 4 ml of liquid M9 medium and grown overnight at 37°C. On the following day, 25 ml of liquid M9 medium containing chloramphenicol, IPTG and 17 mM or 0 mM Glyphosate 25 (Aldrich, 33775-7) were inoculated with 1-2 mls of each overnight culture (in duplicates), the starting OD (at 600 nm) was measured and all the cultures were normalized to start at the same OD. An OD measurement was taken every hour for seven hours. As a control of the bacterial growth, a culture of untransformed *Salmonella* was also inoculated into plain LB medium. In two independent experiments, the clones A₁₇₇I₁₇₈, A₁₇₇V₁₇₈, A₁₇₇L₁₇₈ and I₁₇₇ 30 did not grow in M9 medium, therefore the glyphosate-resistance assays could not be performed on them.

35 6.1.7 ISOLATION AND PURIFICATION OF THE EXPRESSED PROTEIN FROM BACTERIAL CLONES

One milliliter of overnight culture of each of the bacterial clones is inoculated into 100 ml of liquid LB medium containing chloramphenicol. The cells were

allowed to grow at 37°C until they reached an OD of 0.5-0.7 (approximately 3 ½ hours). IPTG was then added to the cultures to a concentration of 1.0 mM. The cells were grown five additional hours. They were then pelleted at 4000 rpm for 20 minutes at 4°C.

The isolation and the purification of the His-tagged proteins were performed
5 following the Qiagen Ni-NTA Protein Purification System. Cell lysates and eluates were
run in duplicates on 12.5% acrylamide gels. One of the gels was silver-stained for
immediate visualization, the second gel was transferred onto Millipore Immobilon-P
membrane, and blocked overnight in 5% milk in TBS-T. The membrane was then exposed
10 to Anti-His primary antibody solution (Amersham Pharmacia biotech, cat# 37-4710),
followed by exposure to Anti-Mouse-IgG secondary antibody solution. (NIF825, from
Amersham Pharmacia biotech ECLWestern blotting analysis system, cat# RPN2108).
Washes and detection reactions were performed according to the manufacturer instructions.
Autoradiograms were developed after 5 minutes exposure.

15 6.2 RESULTS

Cells containing a mutation in the EPSPS gene produced cells that were both
resistant to the herbicide glyphosate and that had a substantially similar growth rate in the
absence or presence of glyphosate, as compared to the wild-type cells, irrespective of the
presence of glyphosate (see Figure 6).

20 It was also demonstrated that the *Arabidopsis* clones containing a mutant
EPSPS gene expressed the mutant protein at substantially the same level as the wild-type
protein (see Figure 7).

25 The invention claimed and described herein is not to be limited in scope by
the specific embodiments, including but not limited to the deposited microorganism
embodiments, herein disclosed since these embodiments are intended as illustrations of
several aspects of the invention. Indeed, various modifications of the invention in addition
to those shown and described herein will become apparent to those skilled in the art from
the foregoing description. Such modifications are also intended to fall within the scope of
the appended claims.

30 A number of references are cited herein, the entire disclosures of which are
incorporated herein, in their entirety, by reference.

WE CLAIM:

1. A non-transgenic herbicide resistant plant, which plant expresses a mutant EPSPS gene product and which plant has substantially normal growth as compared to a 5 plant expressing the wild-type EPSPS gene product.
2. A non-transgenic herbicide resistant plant, which plant expresses a mutant EPSPS gene product, which gene product has substantially the same level of catalytic activity as compared to the wild-type gene product.

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3. The plant according to claim 1 or 2 in which the herbicide is a member of the phosphonomethylglycine family.

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4. The plant according to claim 3 in which the member of the phosphonomethylglycine family is glyphosate.

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5. The plant according to claim 1 or 2 in which the EPSPS gene is mutated at one or more amino acid positions, said positions selected from the group consisting of Leu₁₇₃, Gly₁₇₇, Thr₁₇₈, Ala₁₇₉, Met₁₈₀, Arg₁₈₁, Pro₁₈₂, Ser₉₈, Ser₂₅₅ and Leu₁₉₈ in *Arabidopsis* or at an analogous amino acid residue in an EPSPS paralog.

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6. The plant according to claim 5 in which the positions in the *Zea mays* paralog are selected from the group consisting of Leu₉₇, Gly₁₀₁, Thr₁₀₂, Ala₁₀₃, Met₁₀₄, Arg₁₀₅, Pro₁₀₆, Ser₂₃, Ser₁₇₉ and Leu₁₂₂.

7. The plant according to claim 5 in which the positions in the *Brassica napus* paralog are selected from the group consisting of Leu₁₆₉, Gly₁₇₃, Thr₁₇₄, Ala₁₇₅, Met₁₇₆, Arg₁₇₇, Pro₁₇₈, Ser₉₄, Ser₂₅₁ and Leu₁₉₄.

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8. The plant according to claim 5 in which the positions in the *Petunia hybrida* are selected from the group consisting of Leu₁₆₉, Gly₁₇₃, Thr₁₇₄, Ala₁₇₅, Met₁₇₆, Arg₁₇₇, Pro₁₇₈, Ser₉₄, Ser₂₅₁ and Leu₁₉₄.

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9. The plant according to claim 1 or 2 in which the plant is selected from the group consisting of corn, wheat, rice, barley, soybean, cotton, sugarbeet, oilseed rape,

canola, flax, sunflower, potato, tobacco, tomato, alfalfa, poplar, pine, eukalyptus, apple, lettuce, peas, lentils, grape and turf grasses.

10. The plant according to claim 5 in which the mutated gene results in one
5 or more of the following amino acid substitutions in the EPSPS enzyme in comparison with
the wild-type sequence:

- (i) Leu₁₇₃ - Phe
- (ii) Gly₁₇₇ - Ala or Ile
- (iii) Thr₁₇₈ - Ile or Val or Leu
- 10 (iv) Ala₁₇₉ - Gly
- (v) Met₁₈₀ - Cys
- (vi) Arg₁₈₁ - Leu or Ser
- (vii) Pro₁₈₂ - Leu or Ser
- (viii) Ser₉₈ -Asp
- 15 (ix) Ser₂₅₅ -Ala
- (x) Leu₁₉₈ -Lys.

11. The plant according to claim 6 in which the mutated gene results in one
or more of the following amino acid substitutions in the EPSPS enzyme in comparison with
20 the wild-type sequence:

- (i) Leu₉₇ - Phe
- (ii) Gly₁₀₁ - Ala or Ile
- (iii) Thr₁₀₂ - Ile or Val or Leu
- (iv) Ala₁₀₃ - Gly
- 25 (v) Met₁₀₄ - Cys
- (vi) Arg₁₀₅ - Leu or Ser
- (vii) Pro₁₀₆ - Leu or Ser
- (viii) Ser₂₃ -Asp
- (ix) Ser₁₇₉ -Ala
- 30 (x) Leu₁₂₂ -Lys.

12. The plant according to claim 7 in which the mutated gene results in one
or more of the following amino acid substitutions in the EPSPS enzyme in comparison with
the wild-type sequence:

- 35 (i) Leu₁₆₉ - Phe
- (ii) Gly₁₇₃ - Ala or Ile

(iii) Thr_{174} - Ile or Val or Leu
(iv) Ala_{175} - Gly
(v) Met_{176} - Cys
(vi) Arg_{177} - Leu or Ser
5 (vii) Pro_{178} - Leu or Ser
(viii) Ser_{94} -Asp
(ix) Ser_{251} -Ala
(x) Leu_{194} -Lys.

10 13. The plant according to claim 8 in which the mutated gene results in one or more of the following amino acid substitutions in the EPSPS enzyme in comparison with the wild-type sequence:

(i) Leu_{169} - Phe
(ii) Gly_{173} - Ala or Ile
15 (iii) Thr_{174} - Ile or Val or Leu
(iv) Ala_{175} - Gly
(v) Met_{176} - Cys
(vi) Arg_{177} - Leu or Ser
(vii) Pro_{178} - Leu or Ser
20 (viii) Ser_{94} -Asp
(ix) Ser_{251} -Ala
(x) Leu_{194} -Lys.

25 14. A method for producing a non-transgenic herbicide resistant or tolerant plant comprising

a. introducing into a plant cell a recombinagenic oligonucleobase to produce a mutant EPSPS gene; and
b. identifying a cell having a mutated EPSPS gene, which cell has substantially normal growth as compared to a corresponding wild-type plant cell.

30 15. A method for producing a non-transgenic herbicide resistant or tolerant plant comprising

a. introducing into a plant cell a recombinagenic oligonucleobase to produce a mutant EPSPS gene; and

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b. identifying a cell having a mutated EPSPS gene, which encoded mutant EPSPS protein has substantially the same catalytic activity as compared to a corresponding wild-type EPSPS protein.

5 16. The method according to claim 14 or 15 in which the recombinagenic oligonucleobase is a mixed duplex nucleotide or a SSMOV.

10 17. The method according to claim 16 in which the mixed duplex nucleotide contains a first homologous region which has a sequence identical to the sequence of at least 6 base pairs of the first fragment of the target EPSPS gene and a second homologous region which has a sequence identical to the sequence of at least 6 based pairs of a second fragment of the target EPSPS gene, and an intervening region which contains at least one nucleobase heterologous to the target EPSPS gene, which intervening region connects the first and second homologous region.

15 18. The method according to claim 14 or 15 in which the recombinagenic oligonucleobase is introduced by electroporation.

20 19. The method according to claim 14 or 15 which the mutant EPSPS gene is mutated at one or more amino acid positions, said positions selected from the group consisting of Leu₁₇₃, Gly₁₇₇, Thr₁₇₈, Ala₁₇₉, Met₁₈₀, Arg₁₈₁, Pro₁₈₂, Ser₉₈, Ser₂₅₅ and Leu₁₉₈ in *Arabidopsis* or at an analogous amino acid residue in an EPSPS paralog.

25 20. The plant according to claim 19 in which the positions in the *Zea mays* paralog are selected from the group consisting of Leu₉₇, Gly₁₀₁, Thr₁₀₂, Ala₁₀₃, Met₁₀₄, Arg₁₀₅, Pro₁₀₆, Ser₂₃, Ser₁₇₉ and Leu₁₂₂.

30 21. The plant according to claim 19 in which the positions in the *Brassica napus* paralog are selected from the group consisting of Leu₁₆₉, Gly₁₇₃, Thr₁₇₄, Ala₁₇₅, Met₁₇₆, Arg₁₇₇, Pro₁₇₈, Ser₉₄, Ser₂₅₁ and Leu₁₉₄.

22. The plant according to claim 19 in which the positions in the *Petunia hybrida* are selected from the group consisting of Leu₁₆₉, Gly₁₇₃, Thr₁₇₄, Ala₁₇₅, Met₁₇₆, Arg₁₇₇, Pro₁₇₈, Ser₉₄, Ser₂₅₁ and Leu₁₉₄.

23. The plant according to claim 14 or 15 in which the plant is selected from the group consisting of corn, wheat, rice, barley, soybean, cotton, sugarbeet, oilseed rape, canola, flax, sunflower, potato, tobacco, tomato, alfalfa, poplar, pine, eukalyptus, apple, lettuce, peas, lentils, grape, turf grasses and *Brassica* sp.

5

24. An isolated mutant EPSPS protein comprising the amino acid sequence depicted in SEQ ID NO:2, in which amino acid position Leu₁₇₃ is replaced with Phe, Gly₁₇₇ is replaced with Ala or Ile, Thr₁₇₈ is replaced with Ile or Val or Leu, Ala₁₇₉ is replaced with Gly, Met₁₈₀ is replaced with Cys, Arg₁₈₁ is replaced with Leu or Ser, Pro₁₈₂ is replaced with Leu or Ser, Ser₉₈ is replaced with Asp, Ser₂₅₅ is replaced with Ala or Leu₁₉₈ is replaced with Lys, which mutant EPSPS protein has increased resistance or tolerance to a herbicide, which herbicide is a member of the phosphonomethylglycine family.

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ABSTRACT

The present invention relates to the production of a non-transgenic plant resistant or tolerant to a herbicide of the phosphonomethylglycine family, *e.g.*, glyphosate. The present invention also relates to the use of a recombinagenic oligonucleobase to make a desired mutation in the chromosomal or episomal sequences of a plant in the gene encoding for 5-enol pyruvylshikimate-3-phosphate synthase (EPSPS). The mutated protein, which substantially maintains the catalytic activity of the wild-type protein, allows for increased resistance or tolerance of the plant to a herbicide of the phosphonomethylglycine family, and allows for the substantially normal growth or development of the plant, its organs, tissues or cells as compared to the wild-type plant irrespective of the presence or absence of the herbicide. The present invention also relates to a non-transgenic plant cell in which the EPSPS gene has been mutated, a non-transgenic plant regenerated therefrom, as well as a plant resulting from a cross using a regenerated non-transgenic plant having a mutated EPSPS gene.

DNA sequence:

cccttcatgttttttagaaacccattatcttcttagggcccaattgaaaacccacatttcacctaaccac
caaaggcttgcacatgttgcacgtgaaacaccaaactaacacgtgtcatactgccagtgttatgataaatgctcataccat
accagagtcatagatgtttttgggtggtaaagattgcggatgccttcttcatttcaccaactccctccaaaccc
aacaatgttatattagcaaagccgccaagtgtaaacgaaaagttataaatttcatttctgtatcttacgtaaattg
gagaaatcaatccccatctcgattgtcaattgaagttctccg
[transit peptide start]
ATGGCGCAAGTTAGCAGAATCTGCAATGGTGTGAGAACCCATCTTATCTCCAATCTCTGAAATCCAGTCACGCAA
ATCTCCCTTATCGGTTCTCTGAAGACGCAGCAGCATCCACGGCTTATCCGATTCGTCGTCGGGGATTGAAGAAGA
GTGGGATGACGTTAATTGGCTCTGAGCTCGTCTTAAGGTATGTCCTGTGTTCCACGGCGGAG
[mature peptide starts]
AAAGCGTCGGAGATTGTACTTCACCCATTAGAGAAATCTCGGTCTTATTAAGCTTCTGGCTCCAAGTCTATCAA
TCGGATCTGCTTCTCGTCTGTGAGGTATATTCACCTCGTTCTGCTCTGTAAATCTGAACCTAGATTAT
AAAGATTGATACTTTACCATTTGCTGTGGTTTATAGGGAAACAACGTGAGTGGACAACCTGTTGAATAGCGATGACATC
AATTACATGCTGATCGTTGAAGAGATTGGAACTTAATGTGAAACTGACAGTGAACAAATAATCGTGTAGTTGAAGG
ATGTGGCGGGATATTCCACAGCTTCCATAGATCAGAAAGGTGATATCGAACCTTACCTCGTAATGCAGGAACAGCAATGC
GTCCACTTACCGCTGCGTCACTGCTGAGGTGAAACCGCAAGGTGATGTTGAAGGAGTTGATGCTTCTGGTATTGATG
TTAAGGAATGGAGCTTTGTTGATGCTTATGATCCATTATCCAGTATGTGCTGATGGGGTGCCTCGTATGAGAG
AAAGACCTATAGGGGATTGGTGTGGCTTAAGCAGCTGGTGTGATGTTGAATGTACTCTGGAACACTGCCCCT
CCTGTTCGTGTCAACGCTAATGGTGGCCTCCGGTGGAAAGGTAGATCTGCAAATGGCATGTGAATATGTAATCTCG
TTCCTTACTCTATGAACACTTGCAAGAAATGTGTTCATCATAGCCTTAGCTGACAAGATTCACTTAACTACTC
TCAACGGATGGATCCTAAATAGAACATCGGATTGGTGTGATTGTTCTGTTCTGATTACCGTTCTGTTGATGATTCT
TGATTAACAATTAGGAGACATGTTATGCACTTGAGGTGAAGCTTCTGATCAATTAGTAGTCAGTACTGCTCT
GCTCATGTCCTCTAGCTTGGAGACGTCGAGATTGCGATAATTCTGTTCCATATGTTGAAA
TGACATTGAAGTTAGTGAACGTTGGGGTAGTGTGCGAGCATAGTGATAGCTGGGATGTTCTGTCAAGGGCGGG
AAAAAATACAAGTAGGAGTTATTCTTCTTCTGAATCACATCCCTAGCTGACAATATAATGACTAAAGG
TGAATGATTCAAGGCTCCGGTAATCGTATGAGAGGTGATGCTTCTAGTGATGTTATTCTGGCTGGTGTGCC
TTACCGGTGAAACTGTACAGTCGAAGGTTGTGAACTACCAAGCTGCAAGGTAAATTGTCACACTGAATCATCGACGAG
GCTGTTAAGTTATAGTGAACATTGCTCTAGGTCAAAGTTCATCTTGTATGACAAGGTTGATATAACATATTGCAAGATT
TAAGCTCAATTGGTGTGATGAATCTAGGGAGATGAAAAATTGGGGAGGTCTTGAGAAAATGGGATGTAAGTGTCC
TGGACAGAGAACAGTGTGACTGTGACAGGACCACTAGAGATGCTTTGGAATGAGACACTTGCGGGCTATTGATGTC
CATGAACAAATGCCTGATGTTAGCCATGACCTTGCCGTCGTTCTCTTGCTGACGGTCCAACCACCATAGAGATG
GTAAGTAAAGCTCTCTTATAATTAAAGGTTCTCAATTGATGATCACTTAATTGTTGGTTAATATAGTGGCT
AGCTGGAGAGTAAAGGAGACAGAAAGGATGATTGCCATTGCAAGAGCTAGAAAAGTAAGAGATTCTTATCTCT
TTCTGTCCTTGACAGTGTCTTCAAGTAAATTGTTGTTGTTGAGCTGGGAGCTACAGTGG
AGAAGGTTCAAGATTGGTGTGATAACTCCGCCAAAAGGTGAAACGGCAGAGATTGATGATACATATGATGATCATAGAA
TGGCAATGGCATTCTCTTGCAGCTTGTGCTGATGTTCCATCACCATACAGACTCTGGTTGCACCAGGAAACCTTC
CCGACTACTTCAAGTACTGAAAGAACATCACAAAGCACTAAacaataaaactctgtttttcttctgatccaagctt

FIG. 1A

Protein sequence:

MAQVSRICNGVQNPSLISNLSKSSQRKSPLSVSLKTQQHPRAYPISSWGLKKSGMTLIGSELRPLKVMSSVSTAE
KASEIVLQPIREISGLIKLPGSKSLSNRILLALSEGTTVVDNLLNSDDINYMALDKRLGLNVETDSENNRAVV
EGCGGIFPASIDSKSDIELYLGNAGTAMRPLTAAVTAAGGNASYVLDGVPRMRERPIGDLVVGLKQLGADVECTLG
TNCPPVRVNANGGLPGGKVLSGSISSQYLTALIMSAPALGDVEIEIVDKLISVPYVEMTLKLMERFGSVEHSD
SWDRFFVKGGQKYKSPGNAYVEGDASSACYFLAGAAITGETVTVEGCGTTSLQGDVKFAEVLEKMGCKVSWTENSV
TWTGPPRDAFGMRHLRAIDVNMMNKMPDVAMTLAVVFADGPTTIRDVASWRVKETERMIAICTELRKLGATVEEG
SDYCVITPPKKVKTAEIDTYDDHRMAMAFSLAACADVPITINDSGCTRKTFPDYFQVLERITKH

FIG. 1B

Arabidopsis thaliana wild type sequence:

Position 173 174 175 176 177 178 179 180 181 182 183
L G N A G T A M R P L
CTC GGT AAT GCA **GGA ACA** GCA ATG CGT **CCA** CTT

Arabidopsis thaliana mutant sequences:

Name	CTC GGT AAT GCA GCA ACA GCA ATG CGT CCA CTT L G N A A T A M R P L
A₁₇₇	CTC GGT AAT GCA GCA ACA GCA ATG CGT CCA CTT L G N A A T A M R P L
I₁₇₈	CTC GGT AAT GCA GGA ATA GCA ATG CGT CCA CTT L G N A G I A M R P L
A₁₇₇I₁₇₈	CTC GGT AAT GCA GCA ATA GCA ATG CGT CCA CTT L G N A A I A M R P L
I₁₇₈S₁₈₂	CTC GGT AAT GCA GGA ATA GCA ATG CGT TCA CTT L G N A G I A M R S L
A₁₇₇S₁₈₂	CTC GGT AAT GCA GCA ACA GCA ATG CGT TCA CTT L G N A A T A M R S L
A₁₇₇I₁₇₈S₁₈₂	CTC GGT AAT GCA GCA ATA GCA ATG CGT TCA CTT L G N A A I A M R S L
V₁₇₈S₁₈₂	CTC GGT AAT GCA GGA GTA GCA ATG CGT TCA CTT L G N A G V A M R S L
L₁₇₈S₁₈₂	CTC GGT AAT GCA GGA TTA GCA ATG CGT TCA CTT L G N A G L A M R S L
A₁₇₇V₁₇₈	CTC GGT AAT GCA GCA GTA GCA ATG CGT CCA CTT L G N A A V A M R P L
A₁₇₇L₁₇₈	CTC GGT AAT GCA GCA TTA GCA ATG CGT CCA CTT L G N A A L A M R P L

FIG. 2

FIG. 3A

FIG. 3B

FIG. 3C

FIG. 4

10 20 30 40 50 60 70 80 90 100
 1 MAQVSRLQGIVQNP-SLISNESSQRQSKPSLVSILK1QDHPRAPISSWGLKSGATLIGSELR
 1 MAQSSRLQGIVQNPCT1SMLSKSNQNKSPESVLSLKTHQ
 1 MAQINNADGIQTL-NPNSNFHKPQVKPSSFLVGSKK
 1 AG
 110 120 130 140 150 160 170 180 190 200 210
 104 RILLLAALSEGTTVVDNLLNSDDINMILDALKRLGLMWETDSENNRAVVEGGGLIPASLDSKSDILEYLGNAGTAMRPLTAVYAGGNA
 100 RILLLAALSEGTTVVDNLLNSDDINMILDALKRLGLMWETDSENNRAVVEGGGLIPASLDSKSDILEYLGNAGTAMRPLTAVYAGGNA
 100 RILLLAALSEGTTVVDNLLNSDDINMILDALKRLGLMWETDSENNRAVVEGGGLIPASLDSKSDILEYLGNAGTAMRPLTAVYAGGNA
 100 RILLLAALSEGTTVVDNLLNSDDINMILDALKRLGLMWETDSENNRAVVEGGGLIPASLDSKSDILEYLGNAGTAMRPLTAVYAGGNA
 29 RILLLAALSEGTTVVDNLLNSDDINMILDALKRLGLMWETDSENNRAVVEGGGLIPASLDSKSDILEYLGNAGTAMRPLTAVYAGGNA
 220 230 240 250 260 270 280 290 300 310 320
 214 VGLKQLGADVECTLGTNCPPVRYNANGGLPGGKVKGSSISQQTLA
 210 VGLKQLGADVECTLGTNCPPVRYNANGGLPGGKVKGSSISQQTLA
 210 DGLXQLGAEVDCEFLERKCPVPRIVSKGGPGGVKU
 138 VGLKQLGADVECTLGTNCPPVRYNGLCPGKVKGSSISQQTLA
 330 340 350 360 370 380 390 400 410 420 430
 234 VYEGDASSACYFLAGAAITGETTVVGGCTSLQGDNKRAEVLERKGCKVSTWNTSVTYTGPPRDATGMLRATDVYNNKMPDVANTL
 320 VYEGDASSACYFLAGAAITGETTVVGGCTSLQGDNKRAEVLERKGCKVSTWNTSVTYTGPPRDATGMLRATDVYNNKMPDVANTL
 320 VYEGDASSACYFLAGAAITGETTVVGGCTSLQGDNKRAEVLERKGCKVSTWNTSVTYTGPPRDATGMLRATDVYNNKMPDVANTL
 248 VYEGDASSACYFLAGAAITGETTVVGGCTSLQGDNKRAEVLERKGCKVSTWNTSVTYTGPPRDATGMLRATDVYNNKMPDVANTL
 440 450 460 470 480 490 500 510 520
 434 KETERMIAICTELRLKGATVEEGSDYCWITPPKKVTKAELDTYDDHMMAMAFSLAACADYEVPTIKDPGCTRKTFPDYFOYLESITKH
 430 KETERMIAICTELRLKGATVEEGSDYCWITPPKKVTKAELDTYDDHMMAMAFSLAACADYEVPTIKDPGCTRKTFPDYFOYLESITKH
 430 KETERMIAICTELRLKGATVEEGSDYCWITPPKKVTKAELDTYDDHMMAMAFSLAACADYEVPTIKDPGCTRKTFPDYFOYLESITKH
 358 KETERMIAICTELRLKGATVEEGSDYCWITPPKKVTKAELDTYDDHMMAMAFSLAACADYEVPTIKDPGCTRKTFPDYFOYLESITKH

Oligo Name Oligo Sequence (5'→3')

ATEPS-A₁₇₇

CGTTTCCACCTGCAGCAGTGACCGCAGCGGTAAGTGGACGCATTGCTGTGCTGCATTACCGAG

ATEPS-AI

CGTTTCCACCTGCAGCAGTGACCGCAGCGGTAAGTGGACGCATTGCTATTGCTGCATTACCGAG

ATEPS-IS

CGTTTCCACCTGCAGCAGTGACCGCAGCGGTAAGTGAACGCATTGCTATTGCTGCATTACCGAG

ATEPS-AS

CGTTTCCACCTGCAGCAGTGACCGCAGCGGTAAGTGAACGCATTGCTGTGCTGCATTACCGAG

ATEPS-AIS

CGTTTCCACCTGCAGCAGTGACCGCAGCGGTAAGTGAACGCATTGCTATTGCTGCATTACCGAG

ATEPS-I₁₇₇

CGTTTCCACCTGCAGCAGTGACCGCAGCGGTAAGTGGACGCATTGCTGTATTGCTGCATTACCGAG

ATEPS-VS

CGTTTCCACCTGCAGCAGTGACCGCAGCGGTAAGTGAACGCATTGCTACTGCTGCATTACCGAG

ATEPS-LS

CGTTTCCACCTGCAGCAGTGACCGCAGCGGTAAGTGAACGCATTGCTAATGCTGCATTACCGAG

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FIG. 5

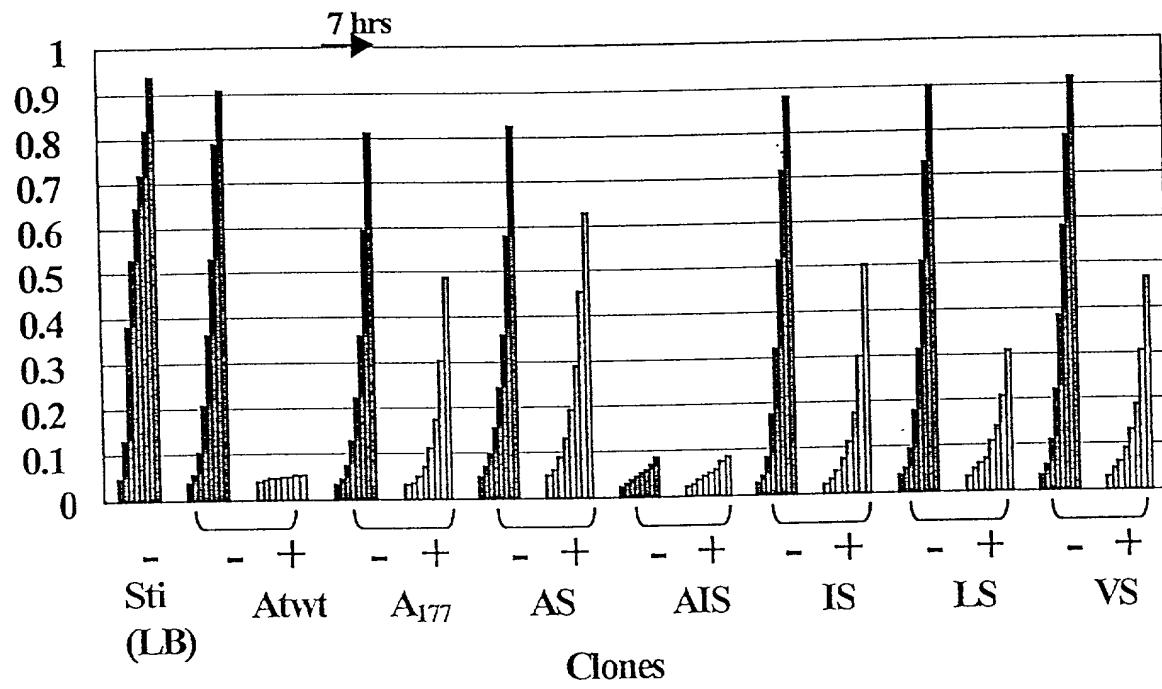


FIG. 6

Arabidopsis clones

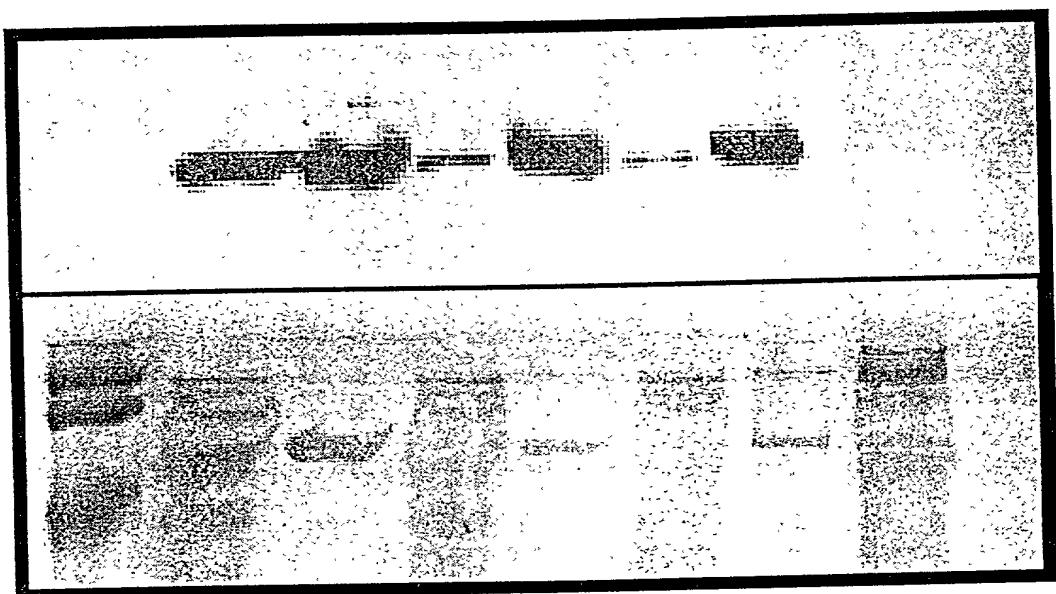
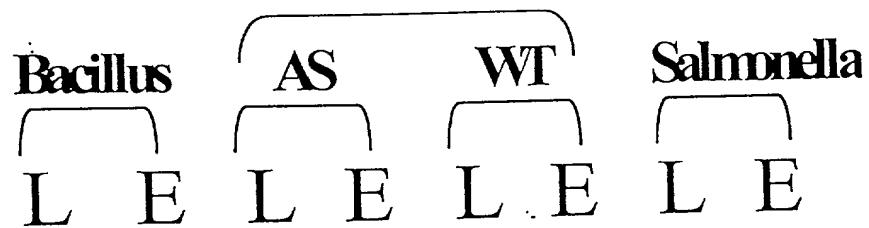


FIG. 7

**DECLARATION
AND POWER OF ATTORNEY**

As a below named inventor, I hereby declare that:

My residence, post office address and citizenship are as stated below at 201 et seq. underneath my name.

I believe I am the original, first and sole inventor if only one name is listed at 201 below, or an original, first and joint inventor if plural names are listed at 201 et seq. below, of the subject matter which is claimed and for which a patent is sought on the invention entitled

NON-TRANSGENIC HERBICIDE RESISTANT PLANTS

and for which a patent application:

is attached hereto and includes amendment(s) filed on *(if applicable)*
 was filed in the United States on as Application No. *(for declaration not accompanying application)*
 with amendment(s) filed on *(if applicable)*
 was filed as PCT international Application No. on and was amended under PCT Article 19 on *(if applicable)*

I hereby state that I have reviewed and understand the contents of the above identified application, including the claims, as amended by any amendment referred to above.

I acknowledge the duty to disclose information known to me to be material to patentability as defined in Title 37, Code of Federal Regulations, §1.56.

I hereby claim foreign priority benefits under Title 35, United States Code, §119(a)-(d) of any foreign application(s) for patent or inventor's certificate listed below and have also identified below any foreign application for patent or inventor's certificate having a filing date before that of the application on which priority is claimed:

EARLIEST FOREIGN APPLICATION(S), IF ANY, FILED PRIOR TO THE FILING DATE OF THE APPLICATION			
APPLICATION NUMBER	COUNTRY	DATE OF FILING (day, month, year)	PRIORITY CLAIMED
			YES <input type="checkbox"/> NO <input type="checkbox"/>
			YES <input type="checkbox"/> NO <input type="checkbox"/>

I hereby claim the benefit under Title 35, United States Code, §119(e) of any United States provisional application(s) listed below.

APPLICATION NUMBER	FILING DATE
60/158,027	October 7, 1999
60/173,564	December 30, 1999

I hereby claim the benefit under Title 35, United States Code, §120 of any United States application(s) listed below and, insofar as the subject matter of each of the claims of this application is not disclosed in the prior United States application in the manner provided by the first paragraph of Title 35, United States Code §112, I acknowledge the duty to disclose information which is material to patentability as defined in Title 37, Code of Federal Regulations, §1.56 which became available between the filing date of the prior application and the national or PCT international filing date of this application:

APPLICATION SERIAL NO.	FILING DATE	STATUS		
		PATENTED	PENDING	ABANDONED

POWER OF ATTORNEY. As a named inventor, I hereby appoint S. Leslie Misrock (Reg. No. 18872), Harry C. Jones, III (Reg. No. 20280), Berj A. Terzian (Reg. No. 20060), David Weild, III (Reg. No. 21094), Jonathan A. Marshall (Reg. No. 24614), Barry D. Rein (Reg. No. 22411), Stanton T. Lawrence, III (Reg. No. 25736), Charles E. McKenney (Reg. No. 22795), Philip T. Shannon (Reg. No. 24278), Francis E. Morris (Reg. No. 24615), Charles E. Miller (Reg. No. 24576), Gidon D. Stern (Reg. No. 27469), John J. Lauter, Jr. (Reg. No. 27814), Brian M. Poissant (Reg. No. 28462), Brian D. Coggio (Reg. No. 27624), Rory J. Radding (Reg. No. 28749), Stephen J. Harbulak (Reg. No. 29166), Donald J. Goodell (Reg. No. 19766), James N. Palik (Reg. No. 25510), Thomas E. Fribel (Reg. No. 29258), Laura A. Coruzzi (Reg. No. 30742), Jennifer Gordon (Reg. No. 30753), Allan A. Fanucci (Reg. No. 30256), Geraldine F. Baldwin (Reg. No. 31232), Victor N. Balancia (Reg. No. 31231), Samuel B. Abrams (Reg. No. 30605), Steven I. Wallach (Reg. No. 35402), Marcia H. Sundeen (Reg. No. 30893), Paul J. Zegger (Reg. No. 33821), Edmond R. Bannon (Reg. No. 32110), Bruce J. Barker (Reg. No. 33291), Adriane M. Antler (Reg. No. 32605), Thomas G. Rowan (Reg. No. 34419), James G. Markey (Reg. No. 31636), Thomas D. Kohler (Reg. No. 32797), Scott D. Stimpson (Reg. No. 33607), Gary S. Williams (Reg. No. 31066), William S. Galliani (Reg. No. 33885), Ann L. Gisolfi (Reg. No. 31956), Todd A. Wagner (Reg. No. 35399), Scott B. Famulant (Reg. No. 35514), Kelly D. Talcott (Reg. No. 39582), Francis D. Cerrito (Reg. No. 38100), Anthony M. Insogna (Reg. No. 35203), Brian M. Rothery (Reg. No. 35340), Brian D. Siff (Reg. No. 35679), and Alan Tenenbaum (Reg. No. 34939), all of Pennie & Edmonds LLP, whose addresses are 1155 Avenue of the Americas, New York, New York 10036, 1667 K Street N.W., Washington, DC 20006 and 3300 Hillview Avenue, Palo Alto, CA 94304, and each of them, my attorneys, to prosecute this application, and to transact all business in the Patent and Trademark Office connected therewith.

SEND CORRESPONDENCE TO:		PENNIE & EDMONDS LLP 1155 Avenue of the Americas New York, NY 10036-2711			DIRECT TELEPHONE CALLS TO: PENNIE & EDMONDS LLP DOCKETING (212) 790-2803	
201	FULL NAME OF INVENTOR	LAST NAME BEETHAM	FIRST NAME PETER	MIDDLE NAME R.		
	RESIDENCE & CITIZENSHIP	CITY CARLSBAD	STATE OR FOREIGN COUNTRY CALIFORNIA	COUNTRY OF CITIZENSHIP AUSTRALIA		
	POST OFFICE ADDRESS	STREET 7128 TANAGER DRIVE	CITY CARLSBAD	STATE OR COUNTRY CALIFORNIA	ZIP CODE 92009	
202	FULL NAME OF INVENTOR	LAST NAME AVISSAR	FIRST NAME PATRICIA	MIDDLE NAME L.		
	RESIDENCE & CITIZENSHIP	CITY EAST BRUNSWICK	STATE OR FOREIGN COUNTRY NEW JERSEY	COUNTRY OF CITIZENSHIP UNITED STATES		
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203	FULL NAME OF INVENTOR	LAST NAME WALKER	FIRST NAME KEITH	MIDDLE NAME A.		
	RESIDENCE & CITIZENSHIP	CITY SAN DIEGO	STATE OR FOREIGN COUNTRY CALIFORNIA	COUNTRY OF CITIZENSHIP UNITED STATES		
	POST OFFICE ADDRESS	STREET 13315 ROXTON CIRCLE	CITY SAN DIEGO	STATE OR COUNTRY CALIFORNIA	ZIP CODE 92130	
204	FULL NAME OF INVENTOR	LAST NAME METZ	FIRST NAME RICHARD	MIDDLE NAME A.		
	RESIDENCE & CITIZENSHIP	CITY LAWRENCEVILLE	STATE OR FOREIGN COUNTRY NEW JERSEY	COUNTRY OF CITIZENSHIP UNITED STATES		
	POST OFFICE ADDRESS	STREET 37 WINTHROP ROAD	CITY LAWRENCEVILLE	STATE OR COUNTRY NEW JERSEY	ZIP CODE 08648	

I HEREBY DECLARE THAT ALL STATEMENTS MADE HEREIN OF MY OWN KNOWLEDGE ARE TRUE AND THAT ALL STATEMENTS MADE ON INFORMATION AND BELIEF ARE BELIEVED TO BE TRUE; AND FURTHER THAT THESE STATEMENTS WERE MADE WITH THE KNOWLEDGE THAT WILLFUL FALSE STATEMENTS AND THE LIKE SO MADE ARE PUNISHABLE BY FINE OR IMPRISONMENT, OR BOTH, UNDER SECTION 1001 OF TITLE 18 OF THE UNITED STATES CODE AND THAT SUCH WILLFUL FALSE STATEMENTS MAY JEOPARDIZE THE VALIDITY OF THE APPLICATION OR ANY PATENT ISSUING THEREON

SIGNATURE OF INVENTOR 201 PETER R. BEETHAM DATE	SIGNATURE OF INVENTOR 202 PATRICIA L. AVISSAR DATE	SIGNATURE OF INVENTOR 203 KEITH A. WALKER DATE
SIGNATURE OF INVENTOR 203 RICHARD A. METZ DATE		

EXPRESS MAIL NO.: EL 501 634 414 US

IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

Application of: Beetham et al.

Application No.: To be assigned

Group Art Unit: To be assigned

Filed: On even date herewith

Examiner: To be assigned

For: NON-TRANSGENIC HERBICIDE
RESISTANT PLANTS

Attorney Docket No.: 7991-086

TRANSMITTAL OF SEQUENCE LISTING UNDER 37 C.F.R. § 1.821

Assistant Commissioner for Patents
Washington, D.C. 20231

Sir:

In accordance with 37 C.F.R. § 1.821, Applicants, in connection with the above-identified patent application, submit herewith a Sequence Listing in paper and computer readable form pursuant to 37 C.F.R. §§ 1.821(c) and (e).

I hereby state that the content of the paper and computer readable copies of the Sequence Listing, submitted in accordance with 37 C.F.R. §§ 1.821(c) and (e), respectively, are the same.

Respectfully submitted,

by Hillel Reuman
REU NO 40,203

Thomas E. Friebel 29,258
Thomas E. Friebel (Reg.No.)

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Enclosures

SEQUENCE LISTING

<110> Beetham, P.
 Avissar, P.
 Walker, K.
 Metz, R.

<120> NON-TRANSGENIC HERBICIDE RESISTANT PLANTS

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<212> DNA
<213> Petunia hybrida

<400> 24

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<211> 1335
<212> DNA
<213> Zea mays

<400> 25

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Val	Ser	Leu	Lys	Thr	His	Gln	Pro	Arg	Ala	Ser	Ser	Trp	Gly	Leu	Lys		
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Lys	Ser	Gly	Thr	Met	Leu	Asn	Gly	Ser	Val	Ile	Arg	Pro	Val	Lys	Val		
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Thr	Ala	Ser	Val	Ser	Thr	Ser	Glu	Lys	Ala	Ser	Glu	Ile	Val	Leu	Gln		
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Pro	Ile	Arg	Glu	Ile	Ser	Gly	Leu	Ile	Lys	Leu	Pro	Gly	Ser	Lys	Ser		
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Leu	Ser	Asn	Arg	Ile	Leu	Leu	Leu	Ala	Ala	Leu	Ser	Glu	Gly	Thr	Thr		
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Val	Val	Asp	Asn	Leu	Leu	Asn	Ser	Asp	Asp	Ile	Asn	Tyr	Met	Leu	Asp		
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Ala	Leu	Lys	Lys	Leu	Gly	Leu	Asn	Val	Glu	Arg	Asp	Ser	Val	Asn	Asn		
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Arg	Ala	Val	Val	Glu	Gly	Cys	Gly	Gly	Ile	Phe	Pro	Ala	Ser	Leu	Asp		
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Ser	Lys	Ser	Asp	Ile	Glu	Leu	Tyr	Leu	Gly	Asn	Ala	Gly	Thr	Ala	Met		
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Arg	Pro	Leu	Thr	Ala	Ala	Val	Thr	Ala	Ala	Gly	Gly	Asn	Ala	Ser	Tyr		
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Val	Leu	Asp	Gly	Val	Pro	Arg	Met	Arg	Glu	Arg	Pro	Ile	Gly	Asp	Leu		
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Gly	Lys	Val	Lys	Leu	Ser	Gly	Ser	Ile	Ser	Ser	Gln	Tyr	Leu	Thr	Ala		
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Leu	Leu	Met	Ala	Ala	Pro	Leu	Ala	Leu	Gly	Asp	Val	Glu	Ile	Glu	Ile		
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Ile	Asp	Lys	Leu	Ile	Ser	Val	Pro	Tyr	Val	Glu	Met	Thr	Leu	Lys	Leu		
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 Ile Thr Gly Glu Thr Val Thr Val Glu Gly Cys Gly Thr Thr Ser Leu
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 Gln Gly Asp Val Lys Phe Ala Glu Val Leu Glu Lys Met Gly Cys Lys
 355 360 365
 Val Ser Trp Thr Glu Asn Ser Val Thr Val Thr Gly Pro Ser Arg Asp
 370 375 380
 Ala Phe Gly Met Arg His Leu Arg Ala Val Asp Val Asn Met Asn Lys
 385 390 395 400
 Met Pro Asp Val Ala Met Thr Leu Ala Val Val Ala Leu Phe Ala Asp
 405 410 415
 Gly Pro Thr Thr Ile Arg Asp Val Ala Ser Trp Arg Val Lys Glu Thr
 420 425 430
 Glu Arg Met Ile Ala Ile Cys Thr Glu Leu Arg Lys Leu Gly Ala Thr
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 Val Glu Glu Gly Ser Asp Tyr Cys Val Ile Thr Pro Pro Ala Lys Val
 450 455 460
 Lys Pro Ala Glu Ile Asp Thr Tyr Asp Asp His Arg Met Ala Met Ala
 465 470 475 480
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 Gly Cys Thr Arg Lys Thr Phe Pro Asp Tyr Phe Gln Val Leu Glu Ser
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 Ile Thr Lys His
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<210> 27
 <211> 516
 <212> PRT
 <213> Petunia hybrida

<400> 27

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 Val Phe Gly Ser Lys Lys Leu Lys Asn Ser Ala Asn Ser Met Leu Val
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 Leu Lys Lys Asp Ser Ile Phe Met Gln Lys Phe Cys Ser Phe Arg Ile
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 Ser Ala Ser Val Ala Thr Ala Gln Lys Pro Ser Glu Ile Val Leu Gln
 65 70 75 80
 Pro Ile Lys Glu Ile Ser Gly Thr Val Lys Leu Pro Gly Ser Lys Ser
 85 90 95
 Leu Ser Asn Arg Ile Leu Leu Leu Ala Ala Leu Ser Glu Gly Thr Thr
 100 105 110
 Val Val Asp Asn Leu Leu Ser Ser Asp Asp Ile His Tyr Met Leu Gly
 115 120 125
 Ala Leu Lys Thr Leu Gly Leu His Val Glu Glu Asp Ser Ala Asn Gln
 130 135 140
 Arg Ala Val Val Glu Gly Cys Gly Gly Leu Phe Pro Val Gly Lys Glu
 145 150 155 160
 Ser Lys Glu Glu Ile Gln Leu Phe Leu Gly Asn Ala Gly Thr Ala Met
 165 170 175

Arg Pro Leu Thr Ala Ala Val Thr Val Ala Gly Gly Asn Ser Arg Tyr
 180 185 190
 Val Leu Asp Gly Val Pro Arg Met Arg Glu Arg Pro Ile Ser Asp Leu
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 Val Asp Gly Leu Lys Gln Leu Gly Ala Glu Val Asp Cys Phe Leu Gly
 210 215 220
 Thr Lys Cys Pro Pro Val Arg Ile Val Ser Lys Gly Gly Leu Pro Gly
 225 230 235 240
 Gly Lys Val Lys Leu Ser Gly Ser Ile Ser Ser Gln Tyr Leu Thr Ala
 245 250 255
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 260 265 270
 Ile Asp Lys Leu Ile Ser Val Pro Tyr Val Glu Met Thr Leu Lys Leu
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 Met Glu Arg Phe Gly Ile Ser Val Glu His Ser Ser Ser Trp Asp Arg
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 305 310 315 320
 Val Glu Gly Asp Ala Ser Ser Ala Ser Tyr Phe Leu Ala Gly Ala Ala
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 Gln Gly Asp Val Lys Phe Ala Glu Val Leu Glu Lys Met Gly Ala Glu
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 370 375 380
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 385 390 395 400
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 405 410 415
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 420 425 430
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 435 440 445
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 450 455 460
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 465 470 475 480
 Phe Ser Leu Ala Ala Cys Ala Asp Val Pro Val Thr Ile Asn Asp Pro
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 Tyr Ser Lys His
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<210> 28

<211> 444

<212> PRT

<213> Zea mays

<400> 28

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 35 40 45

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 Gly Gly Lys Phe Pro Val Glu Asp Ala Lys Glu Glu Val Gln Leu Phe
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 Leu Gly Asn Ala Gly Thr Ala Met Arg Pro Leu Thr Ala Ala Val Thr
 100 105 110
 Ala Ala Gly Gly Asn Ala Thr Tyr Val Leu Asp Gly Val Pro Arg Met
 115 120 125
 Arg Glu Arg Pro Ile Gly Asp Leu Val Val Gly Leu Lys Gln Leu Gly
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 Asn Gly Ile Gly Gly Leu Pro Gly Gly Lys Val Lys Leu Ser Gly Ser
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 Leu Gly Asp Val Glu Ile Glu Ile Ile Asp Lys Leu Ile Ser Ile Pro
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 260 265 270
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 290 295 300
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 325 330 335
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 355 360 365
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 370 375 380
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 385 390 395 400
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<220>
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